

## Short communication. Physiological effects of *Rhizopogon roseolus* on *Pinus halepensis* seedlings

J. A. Dominguez-Nuñez<sup>1\*</sup>, M. Saiz<sup>1</sup>, C. Calderon and J. A. Saiz de Omeñaca<sup>1</sup>

<sup>1</sup> ETSI Mountains and EUIT Forestry. Polytechnic University of Madrid.  
Avda. Ciudad Universitaria, s/n. 28040 Madrid, Spain

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### Abstract

**Aim of study:** The inoculation of forest seedlings with ectomycorrhizal fungi can improve the morphological and physiological qualities of plants, especially those used for regeneration of arid areas. *Rhizopogon roseolus* is an ectomycorrhizal fungus (ECM) commonly used for reforestation.

In this study, the specific objectives were to know some morphophysiological effects of *Rhizopogon Roseolus* on *Pinus halepensis* seedlings under standard nursery conditions.

**Area of study:** ETSI Montes and EUIT Forestal, Madrid.

**Material and methods:** In nursery, under well watered conditions and peat growing substrates, Aleppo pine seedlings were inoculated with *R. roseolus*. Five months after the inoculations, we examined the growth, water parameters (osmotic potential at full turgor [ $\psi\pi_{full}$ ], osmotic potential at zero turgor [ $\psi\pi_0$ ], and the tissue modulus of elasticity near full turgor [ $E_{max}$ ]), mycorrhizal colonization, and concentration and content of macronutrients in the seedlings. Subsequently, a trial was conducted to assess the root growth potential.

**Main results:** The mycorrhization decreased the height and diameter of mycorrhizal seedlings but increased the root weight and root branching. *R. roseolus* did not cause any significant effect on the regeneration of new roots or on any of the tested hydric parameters, but it did improve N uptake of the seedlings.

**Research highlights:** The mycorrhizal inoculation increased the N uptake.

The mycorrhizal inoculation caused opposite effects on some growth parameters.

**Key words:** osmotic adjustment; elastic adjustment; mineral nutrition; root growth potential; nursery; *Rhizopogon roseolus*; *Pinus halepensis*.

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### Introduction

In reforestation, numerous studies have been conducted with the objective of improving the quality of seedlings produced in nurseries (Caravaca *et al.*, 2005). Among the cultural practices utilized, inoculation with ectomycorrhizal fungi (ECM) and plant growth promoting rhizobacteria (PGPR) has shown promise for improving the quality of seedlings and for increasing their survival in plantations, especially in soils with low microbial activity (Chanway, 1997).

There are many reports of positive mycorrhizal effects on the water uptake and water relations of seedlings during drought (Duddridge *et al.*, 1980; Read and Boyd, 1986). However, it cannot be argued that ectomycorrhizae would always have a beneficial role

in water uptake and water relations; it is still unresolved whether ectomycorrhizae influence the components of water potential or osmotic adjustment in shoots or roots (Lehto and Zwiazek, 2011). The capacity to adjust osmotic potential and increase the elasticity of the cell wall (elastic adjustment) is traditionally associated with the increased ability of plants to withstand water stress.

*Rhizopogon roseolus* is an ectomycorrhizal fungal species very common in *Pinus halepensis* forests; early ECM root colonizers that produce large quantities of rhizomorphs, and enhance water and nutrient uptake of plants (Gobert and Plassard, 2007). The spore inoculum is relatively easy to prepare from field-collected material and is a cheap inoculum with low labor costs (Massicotte *et al.*, 1994). For these reasons, *R. roseolus* is an ectomycorrhizal fungus that has been frequently studied for use in forest nurseries and reforestation in Spain (Parladé *et al.*, 2004; Ortega *et al.*, 2004, Rincon *et al.*, 2005, 2007).

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\* Corresponding author: [josealfonso.dominguez@upm.es](mailto:josealfonso.dominguez@upm.es)  
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The objective of this study is to incorporate new information on the effects of *R. roseolus* on the physiology of *P. halepensis* (Aleppo pine, Pino Carrasco) seedlings grown in nurseries, especially on the water relations of seedlings. For this purpose, Aleppo pine seedlings were inoculated with the mycorrhizal fungus *R. roseolus*. We analyzed the osmotic potential at full turgor ( $\psi\pi_{full}$ ) and zero turgor ( $\psi\pi_0$ ), and the modulus of elasticity near full turgor ( $E_{max}$ ). Additionally, we studied the effect of the inoculation on the growth, nutrient uptake, mycorrhizal colonization, and root growth potential of the seedlings.

## Material and methods

Seeds of *P. halepensis* (Valencia) were collected in 2008 and stored in sealed polyethylene bags at 4°C until planting in Forest Pot 300® containers. A mixture of light and dark peat *Sphagnum* type (sterilized by autoclaving at 120°C for 2 h) plus vermiculite (3:1) were used for the substrate. Before sowing, all seeds were sterilized by immersion in 30% H<sub>2</sub>O<sub>2</sub> for 15 min, which was followed by several rinses with distilled water. In mid-April 2008, *P. halepensis* seeds were sown in 300 cells (6 containers, 50 cells per container). The containers were conducted in a greenhouse in the School of Forestry of Madrid, and the seedlings were watered daily to saturation at temperatures ranging from 20 to 30°C until the inoculations were performed. No fertilizers were applied.

The *Rhizopogon roseolus* inoculum (North Gerona, Spain) was purchased from *Micología Forestal y Aplicada*®. Liquid spore inoculum was prepared by diluting the spore inoculum in distilled water. A concentration of 5–7 × 10<sup>4</sup> spores / ml liquid inoculum was estimated. A two-level (*R. roseolus* inoculation and non-inoculated control), unifactorial design that was distributed randomly in three blocks (1 × 2 × 3) was utilized. The inoculum was applied to the seedlings (1 month old) at two time points (20 ml/injection/plant), that were separated by a period of 15 days (in June and early July). 50% of the plants were injected in the substrata, with a total of 40 ml/plant (2–3 × 10<sup>6</sup> spores/plant). After performing the inoculations, the plants were taken outside to a shade house (E.U.I.T. Forest) and were watered daily to saturation. The average temperature ranged from 6°C to 30°C, and the environmental relative humidity average varied from 20% to 100% during the experimental period.

Nine pressure-volume curves (nine seedlings per treatment) were calculated (Tyree and Hammel, 1972) during November 2008, using shoot xylem pressure potentials (Scholander *et al.*, 1965). From each graph, the following water-relation parameters were obtained: 1) the osmotic potential at full turgor ( $\psi\pi_{full}$ ), 2) the osmotic potential at zero turgor ( $\psi\pi_0$ ), and 3) the modulus of elasticity near full turgor ( $E_{max}$ ) (Cheung *et al.*, 1975). Subsequently, the roots of these inoculated seedlings were later analyzed and confirmed to have the mycorrhizal *Rhizopogon roseolus* fungus (Agerer, 1987-98).

Also, a random sample of 9 plants per treatment was chosen. Shoot heights and basal diameters were recorded, and the mycorrhizal colonization in the roots by the characterization and identification of the mycorrhizae (Agerer, 1987-98) was analyzed. After drying at 70°C for 48 h, the dry weights of shoots and total root mass were measured.

30 whole new plants per treatment were randomly selected, pooled, and the whole plant was finely ground and homogenized. For each group ( $n = 3$ ) of 10 plants, the concentrations and content of N, P, K, Ca and Mg in whole plant tissues were determined, using ICP-Spectroscopy (Perkin-Elmer 400) except N, which was measured by a Leco CHN 600 analyzer.

Subsequently, in early February 2009, nine new plants per treatment were randomly chosen, the height and basal diameter were measured previously, and subsequently, each plant was transplanted into a three-liter prismatic pot that was filled with white perlite. The pots were arranged randomly in the greenhouse and were grown for 21 days under optimal environmental conditions (16–22°C temperature; 95% relative humidity) to facilitate their root growth (Burdett, 1987); the plants were irrigated daily. Finally, each plant was extracted, and the number and total length of new roots (>1 cm) per plant were measured.

One-way ANOVA and Duncan's multiple-range test ( $p < 0.05$ ) were performed for all parameters measured (Statgraphics Plus-Statistical software). For statistical analysis of the root growth potential, the height and diameter were selected as covariates.

## Results and discussion

Despite well watering conditions, *R. roseolus* caused a significant decrease in height (5.7 cm versus 6.3 cm) and diameter (1 mm versus 1.13 mm) of seedlings, but

**Table 1.** Mean values and signification of water-relations parameters, growth parameters, mycorrhizal colonization, nutrient concentration and contents, and root growth potential of *Pinus halepensis* seedlings

Treatment	Non- Inoculated	Inoculated	Pvalue <sup>+</sup>
<b>Water-Relations parameters</b>			
$\psi\pi_{full}$ (MPa)	-1.04 ( $\pm 0.1$ )	-1.13 ( $\pm 0.15$ )	0.6247
$\psi\pi_0$ (MPa)	-1.66 ( $\pm 0.13$ )	-1.63 ( $\pm 0.2$ )	0.8824
$E_{max}$ (MPa)	5.94 ( $\pm 0.81$ )	9.32 ( $\pm 1.62$ )	0.0792
<b>Growth</b>			
Height (cm)	6.31 ( $\pm 0.15$ )	5.69 ( $\pm 0.12$ )	0.0047*
Basal Diameter (mm)	1.13 ( $\pm 0.04$ )	1.01 ( $\pm 0.02$ )	0.0192*
Shoot (g)	0.086 ( $\pm 0.006$ )	0.095 ( $\pm 0.004$ )	0.2363
Root (g)	0.082 ( $\pm 0.008$ )	0.104 ( $\pm 0.005$ )	0.0288*
<b>Mycorrhizal colonization</b>			
<i>Rhizopogon</i> (%)	0	42 ( $\pm 4$ )	0.0000*
E-Strain (%)	0	0.5 ( $\pm 0.2$ )	0.0534
Total root tips <sup>2</sup> (N° g <sup>-1</sup> dw)	1867 ( $\pm 124$ )	2707 ( $\pm 259$ )	0.0100*
Total root tips plant <sup>-1</sup>	149 ( $\pm 12$ )	281 ( $\pm 30$ )	0.0008*
<b>Nutrients concentration</b>			
N (mg g <sup>-1</sup> )	5.89 ( $\pm 0.26$ )	7.34 ( $\pm 0.28$ )	0,0196*
P (mg g <sup>-1</sup> )	0.91 ( $\pm 0.05$ )	0.74 ( $\pm 0.04$ )	0,0582
K (mg g <sup>-1</sup> )	9.71 ( $\pm 0.64$ )	8.14 ( $\pm 0.04$ )	0,0715
Ca (mg g <sup>-1</sup> )	7.13 ( $\pm 1.65$ )	7.38 ( $\pm 0.77$ )	0,8986
Mg (mg g <sup>-1</sup> )	4.11 ( $\pm 0.67$ )	4.23 ( $\pm 0.89$ )	0,9170
<b>Nutrients content</b>			
N (mg plant <sup>-1</sup> )	0.99 ( $\pm 0.04$ )	1.46 ( $\pm 0.06$ )	0.0027*
P (mg plant <sup>-1</sup> )	0.15 ( $\pm 0.01$ )	0.15 ( $\pm 0.01$ )	0,6978
K (mg plant <sup>-1</sup> )	1.63 ( $\pm 0.11$ )	1.62 ( $\pm 0.01$ )	0.9013
Ca (mg plant <sup>-1</sup> )	1.20 ( $\pm 0.28$ )	1.47 ( $\pm 0.15$ )	0.4453
Mg (mg plant <sup>-1</sup> )	0.69 ( $\pm 0.11$ )	0.84 ( $\pm 0.18$ )	0.5126
<b>Root Growth Potential</b>			
N° Roots <sup>3</sup>	1 ( $\pm 0$ )	1 ( $\pm 0$ )	0.4637
Length <sup>3</sup> (cm)	1.01 (0.51)	1.69 ( $\pm 0.51$ )	0.3583

<sup>+</sup> ANOVA one-way (*Rhizopogon* Inoculation Factor) for all the parameters except for Root Growth Potential parameters (ANOVA two-way). \* Indicate significant differences ( $p < 0.05$ ). <sup>1</sup>  $\psi\pi_{full}$ : osmotic potential at full turgor,  $\psi\pi_0$ : osmotic potential at zero turgor and  $E_{max}$ : modulus of elasticity near full turgor. <sup>2</sup> Total root tips: number of total root tips, is referred to as grams of root dry weight. <sup>3</sup> Number and total length of new roots per plant; covariate using the height parameter. Values in parentheses represent the standard error.  $N = 9$  (Water-Relations, Growth, Mycorrhizal colonization and Root Growth Potential parameters);  $N = 3$  (Nutrients parameters).

the root dry weight increased significantly in mycorrhizal seedlings (0.104 g) with respect to non-mycorrhizal seedlings (0.082 g), perhaps by providing phytohormones (Amaranthus and Perry, 1989). The mycorrhizae were morphological identified as *R. roseolus* (42%) and *E-Strain* (0.5%) in the inoculated plants; no other mycorrhizal morphotypes were founded in non-inoculated seedlings; *R. roseolus*

generated an increase in root branching (2707 versus 1867 tips.g<sup>-1</sup>dw) and an increase in the total number of root tips (281 versus 149 tips plant<sup>-1</sup>) of seedlings (Table 1).

A nutrient-poor substrate induced a small growth of all seedlings in general. *R. roseolus* inoculation caused a significant improvement in N uptake (7.34 versus 5.89 mg g<sup>-1</sup>; 1.46 versus 0.99 mg.plant<sup>-1</sup>). By contrast,

the mycorrhization did not cause significant effects on the absorption of the other macronutrients analyzed (Table 1).

Some authors have shown the ability of *R. roseolus* to improve growth and nutrient uptake of seedlings, especially N and P, of pines other than *P. halepensis* under nursery conditions (Rincon *et al.*, 2005). On the contrary, other authors have observed limiting effects (Rincon *et al.*, 2007). Casarin *et al.* (2004) showed in *P. pinaster* that *R. roseolus* facilitates the mobilization of P from soils with low available P. Gobert and Plassard (2007) showed that *R. roseolus* has a positive effect on the absorption of N in *P. pinaster* when the supply is low and fluctuating.

The capacity to adjust osmotic potential and increase the elasticity of the cell wall (elastic adjustment) is related to the resistance of the plants to drought. Through both mechanisms, plants are able to maintain turgor potential, the capacity for growth and photosynthesis, and the ability to tolerate more negative water potential and lower water availability (Villar-Salvador *et al.*, 1997). According to the results obtained in this study, under well watering conditions, *R. roseolus* caused no significant effect on any hydric parameters (Table 1); *R. roseolus* seems to have caused an apparent rigidity of cell walls ( $p$ -value = 0.0792), although results were not significant. Other studies have shown that some mycorrhizal fungi, including *R. roseolus*, are able to improve the water availability of their host plants, especially under drought conditions because of strong development of external mycelium (Parke *et al.*, 1983). Ortega *et al.*, (2004) found that the hydraulic conductance of *R. roseolus* mycorrhizal seedlings was improved in sites under low water availability.

The ability to maintain open stomata and photosynthesis during the early stages of a drought could increase the supply of carbon for growth, particularly new root growth, which requires current photosynthates (van den Driessche, 1987). Moreover, an increased rate of photosynthesis supplies additional carbon for osmotic adjustment, thereby promoting turgor maintenance (Tan *et al.*, 1992) and permitting higher water uptake from drier soils. In this study, the test to analyze the root growth potential showed that the *R. roseolus* did not cause any significant effects on the regeneration of new roots in Aleppo pine seedlings. This lack of response in the test of root regeneration could be related to the absence of significant differences in the osmotic potential analysis of

seedlings under well watering conditions. Additionally, it may have been appropriate to increase the number of plants sampled and the duration of the radical regeneration trial such that the treatment effects would be manifested in the seedlings.

## Conclusions

Under well watering conditions, *R. roseolus* did not provide osmotic adjustment mechanisms and regeneration of new roots in seedlings. However, under these conditions, throughout the first year of nursery cultivation, we demonstrated that *R. roseolus* caused effects such as increased weight and branching of the root, or increases in N uptake (on a substrate poor in macronutrients) in the seedlings, so these effects do not appear to be related to water availability. Further research could study the effect of *R. roseolus* and other ectomycorrhizal fungi under conditions of water stress on the osmotic adjustment, elastic adjustment, root growth potential, and other lesser-known mechanisms that affect the water relationships of their host forest plants.

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## References

- Agerer R, 1987-1998. Colour atlas of ectomycorrhizae. Ed Einhorn-Verlay, Munich, Germany.
- Amaranthus MP, Perry D, 1989. Rapid root tip and mycorrhiza formation and increased survival of Douglas-fir seedlings after soil transfer. *New For* 3: 77-82.
- Burdett AN, 1987. Understanding root growth capacity: theoretical considerations in assessing planting stock quality by means of root growth tests. *Can J Forest Res* 17: 768-775.
- Caravaca F, Alguacil MM, Azcón R, Parladé J, Torres P, Roldán A, 2005. Establishment of two ectomycorrhizal shrub species in a semiarid site after in situ amendment with sugar beet, rock phosphate, and *Aspergillus niger*. *Microbial Ecol* 49: 73-82.

- Casarin V, Plassard C, Hinsinger Ph, Arvieu JC, 2004. Quantification of ectomycorrhizal fungal effects on the bioavailability and mobilization of soil P in the rhizosphere of *Pinus pinaster*. *New Phytol* 163: 177-185.
- Chanway CP, 1997. Inoculation of tree roots with plant growth promoting soil bacteria: An emerging technology for reforestation. *Forest Sci* 43:99-112.
- Cheung YNS, Tyree MT, Dainty J, 1975. Water relations parameters on single leaves obtained in a pressure bomb and some ecological interpretations. *Can J Bot* 53: 1342-1346.
- Duddridge JA, Malibari A, Read DJ, 1980. Structure and function of mycorrhizal rhizomorphs with special reference to their role in water transport. *Nature* 287: 834-836.
- Gobert A, Plassard C, 2007. Kinetics of NO<sup>3-</sup> net fluxes in *Pinus pinaster*, *Rhizopogon roseolus* and their ectomycorrhizal association, as affected by the presence of NO<sup>3-</sup> and NH<sup>4+</sup>. *Plant Cell Environ* 30(10): 1309-1319.
- Lehto T, Zwiazek J, 2011. Ectomycorrhizas and water relations of trees: a review. *Mycorrhiza* 21(2): 71-90.
- Massicote HB, Molina R, Luoma DL, Smith JE, 1994. Biology of the ectomycorrhizal genus *Rhizopogon*. II. Patterns of host-fungus specificity following spore inoculation of diverse hosts grown in monoculture and dual culture. *New Phytol* 126: 677-690.
- Ortega U, Duñabeitia A, Menéndez S, González-Murua C, Majada J, 2004. Effectiveness of mycorrhizal inoculation in the nursery on growth and water relations of *Pinus radiata* in different water regimes. *Tree Physiol* 24: 65-73.
- Parke JL, Linderman RG, Black CH, 1983. The role of ectomycorrhizas in drought tolerance of Douglas-fir seedlings. *New Phytol* 95: 83-95.
- Parladé J, Luque J, Pera J, Rincón A, 2004. Field performance of *Pinus pinea* and *P. halepensis* seedlings inoculated with *Rhizopogon* spp. and outplanted in formerly arable land. *Ann For Sci* 61: 507-514.
- Read DJ, Boyd R, 1986. Water relations of mycorrhizal fungi and their host plants. In: Water, fungi and plants (Ayres PG, Boddy L, eds). Cambridge University Press, Cambridge, UK. pp: 215-240.
- Rincón A, Parladé J, Pera J, 2005. Effects of ectomycorrhizal inoculation and the type of substrate on mycorrhization, growth and nutrition of containerised *Pinus pinea* L. seedlings produced in a commercial nursery. *Ann For Sci* 62: 1-6.
- Rincón A, De Felipe MR, Fernández-Pascual M, 2007. Inoculation of *Pinus halepensis* Mill. with selected ectomycorrhizal fungi improves seedling establishment 2 years after planting in a degraded gypsum soil. *Mycorrhiza* 18: 23-32.
- Scholander PF, Hammel HT, Bradstreet ED, Hemmingsen EA, 1965. Sap pressure in vascular plants. *Science* 148: 339-346.
- Tan W, TJ Blake, Boyle TJB, 1992. Drought tolerance in faster- and slower-growing black spruce (*Picea mariana*) progenies: II. Osmotic adjustment and changes of soluble carbohydrates and amino acids under osmotic stress. *Physiol Plant* 85: 645-651.
- Tyree M, Hammel HT, 1972. The measurement of the turgor pressure and the water relations of plants by the pressure technique. *J Exp Bot* 23: 267-282.
- Van den Driessche R, 1987. Importance of current photosynthate to new root growth in planted conifer seedlings. *Can J For Res* 17: 776-782.
- Villar-Salvador P, Caña L, Peñuelas J, Carrasco I, Domínguez S, Renilla I, 1997. Relaciones hídricas y potencial de formación de raíces en plántulas de *Pinus halepensis* Mill. sometidas a diferentes niveles de endurecimientos por estrés hídrico. In *Monographs of the Spanish Society of Forest Science* 4: 81-92.