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# Post-emergent herbicidal activity of Eucalyptus globulus Labill. essential oil

Actividad herbicida post-emergente del aceite esencial de Eucalyptus globulus Labill

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#### **ABSTRACT**

Weed resistances to synthetic herbicides, as well as consequent health and environmental problems, are important items to find more eco-friendly natural alternatives to weed control. *Eucalyptus globulus* Labill. essential oil has been traditionally used against respiratory troubles as well as an insect repellent due to 1,8-cineole content. Chemical composition of commercial *E. globulus* essential oil and its phytotoxic activity against three common annual weeds (*Portulaca oleracea* L., *Echinochloa crus-galli* (L) Beau. and *Lolium multiflorum* Lam.) has been studied. 28 compounds reaching 99.83% of the total essential oil were identified by gas chromatography-mass spectrometry analysis. The oxygenated monoterpene 1,8-cineole (76.43±0.35%), followed by the monoterpene hydrocarbon  $\alpha$ -pinene (14.64±0.27%) were the main compounds. *E. globulus* essential oil lacks of phytotoxicity against the seed germination of the tested weed, showing significant effect on hypocotyl and radicle elongation of *E. crus-galli* at the highest dose (1  $\mu$ L/mL) assayed and radicle inhibitory effects at all concentrations applied (0.125, 0.25, 0.50 and 1  $\mu$ L/mL) against *L. multiflorum*. *E. globulus* essential oil could be used in the management of *E. crus-galli* due to its post-emergent herbicidal activity.

**KEYWORDS:** Eucalyptus globulus, essential oil, GC-MS, phytotoxic activity.

#### **RESUMEN**

La aparición de resistencias debido al uso de herbicidas sintéticos, así como los consiguientes problemas de salud y medioambientales, son factores importantes en la búsqueda de alternativas naturales respetuosas con el medio ambiente en el control de malas hierbas. El aceite esencial de *Eucalyptus globulus* Labill se ha empleado tradicionalmente en problemas respiratorios y como repelente de insectos por su contenido en 1,8-cineol. Se ha determinado la composición química del aceite esencial comercial de *E. globulus* y su actividad fitotóxica en tres conocidas malas hierbas anuales (*Portulaca oleracea* L., *Echinochloa crus-galli* (L) Beau y *Lolium multiflorum* Lam.). Se identificaron veintiocho compuestos que representan el 99,83 % de la composición total de este, mediante cromatografía de gases - espectrometría de masas. Los componentes mayoritarios fueron el monoterpeno oxigenado 1,8-cineol (76,43 ± 0,35 %) y el monoterpeno hidrocarbonado  $\alpha$ -pineno (14,64 ± 0,27 %). El aceite esencial de *E. globulus* carece de actividad frente a la germinación de semillas de las malas hierbas ensayadas, mostrando efecto inhibitorio signi-

ficativo sobre la elongación del hipocotilo y radícula de E. crus-galli a la dosis más alta (1  $\mu$ L/mL) empleada y sobre el crecimiento de la radícula de L. multiflorum a todas las concentraciones (0,125, 0,25, 0,50 y 1  $\mu$ L/mL) aplicadas. El aceite esencial de E. globulus puede utilizarse en el manejo de E. crus-galli debido a su actividad herbicida postemergente.

PALABRAS CLAVE: Eucalyptus globulus, aceite esencial, CG-EM, actividad fitotóxica.

#### INTRODUCTION

Eucalyptus (Eucalyptus globulus Labill.) is a tree belonging to the Myrtaceae family which leaves are traditionally used in cough and flu disorders by its anticatarrhal, expectorant and antipneumonic properties [1,2]. Its essential oil has been included in a mixture with other volatile oils to relieve muscular aches, arthritis and respiratory troubles [3], being recently corroborated the mucolytic effect of the fluid extract obtained from the leaves of E. globulus together with Borago officinalis L. and Sambucus nigra L. [4]. In addition, its essential oil is also a recognized insecticidal agent commonly used not only as an alternative pediculicide with a 100 % of effectiveness in humans [5] but also as insect repellent against harmful creatures in agriculture, such as housefly (Musca domestica) and Acanthoscelides obtectus [6-8]. In this sense, E. globulus essential oil is being studied for pest control in food production, due to its wide-spectrum antimicrobial activity against storage foodstuff pathogens, like certain bacteria including Escherichia coli and Pseudomonas aeruginosa [9], fungal strains with a dose-dependent fungicidal effect against Aspergillus flavus and A. parasiticus and their aflatoxin production [10] and also against the normal development of Fusarium verticillioides by delaying spore germination causing a reduction in fumonisin production, too [11], as well as against other food spoilage microorganisms, such as yeasts strains (Candida albicans and Sacchromyces cerevisiae) [9]. Regarding this, there is an increased interest in the research of the industrial application of these properties, for instance E. globulus essential oil is incorporated as a natural antimicrobial ingredient in edible films exhibiting its antimicrobial and antioxidant properties and consequently enhancing microbial safety and shelf-life of food [12].

It is interesting to note the pesticide activity of natural compounds and their potential applications, particularly for a sustainable agriculture [13,14] due to the rising agrochemical problematic: the World Health Organization (WHO) warns about synthetic pesticides that have been seen to cause serious public health effects along years as consequence of the presence of considerable levels of pesticide residues in ground and surface water, as well as in food with their subsequently cause of human acute poisonings and even more cancer and other chronic illnesses [15,16]. Furthermore, it is of indispensable consideration the constant emergence of resistances by practically every type of organisms after the extensive use of pesticides making this fact one of the top four environmental problems in the world [15].

According to this, it is popularly known the case of glyphosate, the world's best known herbicide, whose resulting resistances have been described in many worldwide species, like common ragweed (*Ambrosia artemisiifolia* L.) in several row crops of the south-eastern USA following other still unknown mechanisms of action [17], annual ryegrass (*Lolium rigidum* L.) in Australia [18] or barnyard-grass (*Echinochloa crus-galli* (L.) Beauv.) in cotton fields of the midsouthern United States [19].



Consequently, the phytotoxic effect of several essential oils continues to be studied against seed germination and seedling growth of some weeds [20]. For instance, *Citrus aurantiifolia* essential oil has demonstrated herbicidal effect against three agricultural weeds, *Avena fatua*, *Echinochloa crus-galli* and *Phalaris minor*, reducing their germination at ≥0.25-0.50 mg/mL as well as the coleoptile and root growth at ≥0.10-0.50 mg/mL [21]. Thyme (*Thymus vulgaris*), summer savory (*Satureja hortensis*), clove (*Syzgium aromaticum*) and cinnamon (*Cinnamomum zeylanicym*) essential oils have provided phytotoxic results causing electrolyte leakage and cell death of dandelion leaf (*Taraxacum officinale* Weber in Wiggers) [22], showing also *S. hortensis* essential oil nanoemulsion changes on germination, growth and morphophysiological features of *Amaranthus retroflexus* L. and *Chenopodium album* L. [23].

In relation to *E. globulus* essential oil, it is able to exert strong deleterious effects on the germination of *A. retroflexus* and *Portulaca oleracea* L. [24], seed germination and seedling growth of *Parthenium hysterophorus* L. [25] as well as on germination percentage and germination rate, radicle length, plumule length, primary root and pedicle length, and seedling height of *A. blitoides* and *Cynodos dactylon* (L.) Pers., at increasing concentrations [26].

Together the phytotoxic effects it is important to find selective herbicides that only disturb the seed germination and seedling development of weeds, without toxic effects on food crops.

So, the aims of this work are firstly to standardize through gas chromatography-mass spectrometry analysis the chemical composition of the commercial *E. globulus* essential oil in order to assure its main compounds and secondly, to determine their *in vitro* phytotoxic activity against seed germination and seedling growth of *P. oleracea*, a cosmopolitan annual weed of tropical and subtropical climates, *L. multiflorum*, a grass distributed along temperate climates affecting mostly cereals and *E. crus-galli*, an annual plant seriously influencing irrigation crops, especially rice, in order to obtain eco-friendly herbicides.

#### MATERIALS AND METHODS

#### **Essential Oil**

Commercial sample of eucalyptus (*E. globulus* Labill.) (Batch 0065901) essential oil purchased from Guinama Lab. (Valencia, Spain), was stored at 4 °C until chemical analysis and phytotoxic studies were carried out.

#### **Seeds**

Mature seeds of annual weeds of common purslane (*Portulaca oleracea* L.), Italian ryegrass (*Lolium multiflorum* Lam.) and barnyardgrass (*Echinochloa crus-galli* (L.) Beauv.), were purchased from Herbiseed, UK (website: www.herbiseed.com).



# Gas Chromatography-Mass Spectrometry Analysis

GC-MS analysis was carried out with a 5973N Agilent apparatus, equipped with a capillary column (95 dimethylpolysiloxane- 5 % diphenyl), HP-5MS UI (30 m long and 0.25 mm i.d. with 0.25 µm film thickness). The column temperature program was 60 °C during 5 min, with 3 °C/min increases to 180 °C, then 20 °C/min increases to 280 °C, which was maintained for 10 min. The carrier gas was helium at a flow-rate of 1 mL/min. Split mode injection (ratio 1:30) was employed. Mass spectra were taken over the m/z 30-500 range with an ionizing voltage of 70 eV.

#### Identification

The individual compounds were identified by mass spectra and their identity was confirmed by comparison of their Kovat's retention index calculated using standard hydrocarbons relative to  $C_8$ - $C_{32}$  n-alkanes, and mass spectra with reference samples or with data already available in the NIST 2005 Mass Spectral library and in the literature [27].

## **Herbicidal Activity**

Sets of 20 seeds each with five replicates per treatment were homogenously distributed in Petri dishes (9 cm diameter) between two layers of filter paper (Whatman No.1) moistened with 4 mL of distilled water and with 0 (control), 0.125, 0.250, 0.5, and  $1\mu L/mL$  of *E. globulus* essential oil. Petri dishes were sealed with parafilm and incubated in a germination chamber Equitec EGCS 301 3SHR model, according to previous assays [28] alternating  $30.0 \pm 0.1$  °C 16 h in light and  $20.0 \pm 0.1$  °C 8 h in dark and with (*E. crus-galli*) and without (*P. oleracea*, *L. multiflorum*) humidity. To evaluate the herbicidal activity of the essential oil, the number of germinated seeds was counted and compared with those of untreated seedlings. Emergence of the radicle ( $\ge 1$  mm) was used as an index of germination and seedling length (hypocotyl and/or radicle) data was recorder after 3, 5, 7, 10 and 14 days in each replicate.

## **Statistical Analysis**

Experiments were made with five replicates. Resulting data were subjected to one-way analysis of variance with IBM SPSS statistics 22 software. Tukey's *post hoc* test was used when variances remained homogeneous (Levene's test) and T3 Dunnett's *post hoc* one was employed if not, assuming equal variances. Differences were considered to be significant at  $p \le 0.05$ .



#### **RESULTS AND DISCUSSION**

# Chemical Composition of *E. globulus* Essential Oil

Twenty-eight compounds reaching 99.83 % of the total commercial *E. globulus* essential oil were identified by gas chromatography-mass spectrometry analysis. Compounds are clustered (Table 1) in homologous series of monoterpene hydrocarbons, oxygenated monoterpenes, sesquiterpene hydrocarbons, and oxygenated sesquiterpenes and listed according to Kovat's retention index calculated in GC on apolar HP-5MS column.

Highest quantities of monoterpene compounds (98.78 %) were found in *E. globulus* essential oil. Twelve monoterpene hydrocarbons (21.93±0.08 %) and seven oxygenated monoterpenes (76.85±0.13 %) constituted the monoterpene fraction. The oxygenated monoterpene 1,8-cineole (76.43±0.35 %) followed by the monoterpene hydrocarbon α-pinene (14.64±0.27 %) were the main compounds. Among the monoterpene hydrocarbons, also large quantities of β-pinene (3.26±0.03 %) and γ-terpinene (2.00±0.02 %) were found.

 $\beta$ -Caryophyllene and longifolene with 0.51 $\pm$ 0.01 % and 0.30 $\pm$ 0.00 %, respectively, were the principal components among the sesquiterpene hydrocarbons. No higher percentages than 0.1 % were found among the other sesquiterpene hydrocarbons identified.

Finally, caryophyllene oxide  $(0.04\pm0.00 \%)$  was the only oxygenated sesquiterpene found in *E. globulus* essential oil here analyzed (Table 1).

The results obtained were similar to recent research [29] with 1,8-cineole (76.6 %) and  $\alpha$ -pinene (12.9 %) as the main compounds of *E. globulus* essential oil and different from those obtained from samples of Algeria [30] in which 1,8-cineole is the main compound (55.3 %) followed by spathulenol (7.4 %) and  $\alpha$ -terpineol (5.5 %).

*Eucalyptus* spp. are characterized by a great variability of the main compounds in their essential oils. Thus, in samples collected in Pakistan, large amount of citronellal (22.3 %) and citronellol (20.0 %) have been found in *E. citriodora* essential oil. Limonene (14.3 %) and terpinen-4-ol (10.2 %) were the main compounds in *E. crebra* essential oil; 1,8-cineole (15.2 %) and α-pinene (12.1 %) in *E. tereticornis* essential oil, while linalool (17.0 %), followed by 1,8-cineole (16.1 %), were the principal components in *E. camaldulensis* [31]. However, *E. camaldulensis* is also characterized by a high content of spathulenol (41.46 %) and *p*-cymene (21.92 %) in samples collected in Valencia (Spain) [32]. The different chemical composition may be attributed to geographical, environmental and climatic differences [31].



Table 1. Chemical composition of commercial E. globulus essential oil

RT	RI	Compound	E. globulus Peak area (%)
	Monoter	pene hydrocarbons	21.93±0.08
6.58	925	α-Thujene	$0.01 \pm 0.00$
6.88	933	α-Pinene	14.64±0.27
7.36	936	Camphene	$0.18 \pm 0.00$
7.58	952	Thuja-2,4(10)-diene	$0.01 \pm 0.00$
8.48	973	β-Pinene	$3.26 \pm 0.03$
9.11	987	Myrcene	$0.77 \pm 0.01$
9.65	998	α-Phellandrene	$0.58\pm0.01$
10.20	1012	α-Terpinene	$0.14\pm0.01$
11.25	1037	(Z)-β-Ocimene	$0.11 \pm 0.01$
11.68	1047	( <i>E</i> )- β-Ocimene	$0.03 \pm 0.00$
12.13	1056	γ-Terpinene	$2.00\pm0.02$
13.46	1083	Terpinolene	$0.22 \pm 0.01$
	Oxygena	uted monoterpenes	76.85±0.13
11.10	1033	1,8-Cineole	76.43±0.35
13.87	1091	Linalool	$0.02\pm0.01$
14.04	1094	α-Pinene oxide	$0.01\pm0.00$
17.59	1171	Terpinen-4-ol	$0.04 \pm 0.01$
18.01	1479	<i>p</i> -Cymen-8-ol	$0.01 \pm 0.01$
18.24	1184	α-Terpineol	$0.31 \pm 0.02$
22.62	1278	Bornyl acetate	$0.03\pm0.00$
	Sesquiter	pene hydrocarbons	1.01±0.00
25.38	1341	α-Cubebene	$0.05 \pm 0.00$
26.21	1360	Longicyclene	$0.01\pm0.00$
27.71	1393	Longifolene	$0.30\pm0.00$
28.34	1408	β-Caryophyllene	0.51±0.01
29.74	1443	α-Humulene	$0.06 \pm 0.00$
30.72	1467	γ-Muurolene	$0.01 \pm 0.01$
31.68	1490	α-Muurolene	$0.01 \pm 0.00$
32.60	1513	δ-Cadinene	$0.06 \pm 0.00$
	Oxygena	ated sesquiterpenes	0.04±0.00
34.85	1571	Caryophyllene oxide	$0.04 \pm 0.00$
		TOTAL	99.83±0.21

RI, retention index relative to  $C_8$ - $C_{32}$  *n*-alkane on HP-5MS column; values are mean  $\pm$  standard deviation of three samples.

1,8-Cineole, the main constituent of the essential oil from *E. globulus* and other species, is a well-known wide-spectrum antibacterial agent [7]. Synergic effects against antibiotic-resistant pathogens have been observed when combined with the sesquiterpene hydrocarbon aromadendrene, the main



compound of the essential oil from the fruits of *E. globulus* [33], and also with chlorhexidine digluconate, against methicillin-resistant *Staphyloccocus aureus* in planktonic and biofilm cultures [34,35].

On the other hand, its combination with  $\alpha$ -pinene, the second main component of *E. globulus* essential oil analysed here, provides beneficial effects against cellular oxidative stress, preventing reactive oxygen species-induced damage involved in the pathogenesis of several neurodegenerative disorders such as Alzheimer's disease [36].

# Seed Germination and Seedling Growth Inhibition of *P. oleracea*, *L. multiflorum* and *E. crus-galli*, by *E. globulus* Essential Oil

The phytotoxic effect of *E. globulus* essential oil against seed germination and seedling growth of three well-known weeds, *P. oleracea*, *E. crus-galli* and *L. multiflorum*, is shown in Table 2 and Figures 1-3.

*E. globulus* essential oil had no effect against *P. oleracea*, *E. crus-galli* and *L. multiflorum* seed germination. No significant differences were found between control and all doses (0.125, 0.25, 0.5 and 1  $\mu$ L/mL) assayed (Table 2).

Table 2. In vitro effects of E. globulus essential oil against P. oleracea, L. multiflorum
and E. crus-galli seed germination and seedling growth

Commented (I/mI)	P. oleracea				
Concentration (μL/mL) —	Germination	Hypocotyl growth	Radicle growth		
Control	67.00±4.06 a	2.40±0.34 a	1.50±0.28 a		
0.125	70.00±6.12 a	2.33±0.43 a	1.29±0.16 a		
0.25	64.00±2.45 a	2.08±0.13 a	1.32±0.19 a		
0.5	66.00±4.85 a	1.83±0.35 a	1.50±0.33 a		
1	70.00±1.58 a	1.83±0.19 a	1.81±0.22 a		
Concentration (µL/mL)		E. crus-galli			
	Germination	Hypocotyl growth	Radicle growth		
Control	74.00±3.32 a	22.97±1.75 a	18.92±1.26 a		
0.125	87.00±2.55 a	22.95±1.20 a	18.34±1.09 a,b		
0.25	73.00±2.55 a	17.76±1.09 a,b	17.70±0.73 a,b		
0.5	74.00±3.32 a	17.92±1.83 a,b	13.45±0.82 a,b		
1	78.00±5.39 a	15.81±2.14 b	15.38±1.78 b		
Concentration (µL/mL)		L. multiflorum			
	Germination	Hypocotyl growth	Radicle growth		
Control	67.00±2.00 a	29.18±1.26 a	32.92±2.63 a		
0.125	55.00±2.74 a	24.46±0.55 a	19.35±1.22 b		
0.25	63.00±3.39 a	25.08±1.55 a	18.01±0.78 b		
0.5	63.00±5.39 a	24.77±1.97 a	13.95±1.68 b		
1	63.00±3.00 a	24.60±0.61 a	15.49±1.19 b		

<sup>&</sup>lt;sup>a</sup> Values are mean of five replications ± error deviation after 14 days of incubation. Means followed by different letters in the same column indicate that are significantly different at p>0.05 according to T3 Dunnet and Tukey tests.



According to seedling growth, no significant differences in the seedling development (hypocotyl and radicle) of *P. oleracea* were observed after the application of *E. globulus* essential oil at all the concentrations assayed in comparison to control (Table 2, Figure 1).

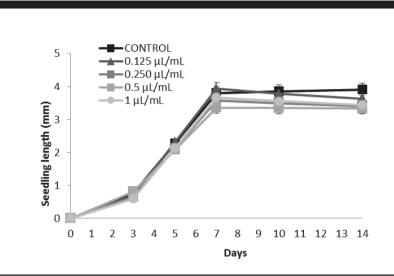


Figure 1. *P. oleracea* seedling growth with *E. globulus* essential oil. Control and treated with *E. globulus* essential oil at 0.125, 0.25, 0.5 and 1 μL/mL.

However, both hypocotyl and radicle of *E. crus-galli* were significantly inhibited at the highest dose tried (1  $\mu$ L/mL) reaching 31.17 and 18.71 % of growth reduction with respect to control (Table 2, Figure 2).

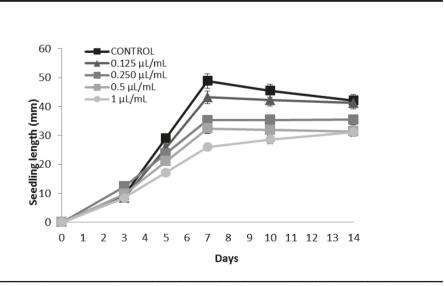


Figure 2. *E. crus-galli* growth with *E. globulus* essential oil. Control and treated with *E. globulus* essential oil at 0.125, 0.25, 0.5 and 1  $\mu$ L/mL.



Regarding *L. multiflorum* evolution, although no significant differences were observed in its hypocotyl growth (Table 2), the radicle development was considerably reduced between 41.22-52.95 % without differences between all concentrations (0.125, 0.25, 0.50 and 1  $\mu$ L/mL) applied (Table 2, Figure 3).

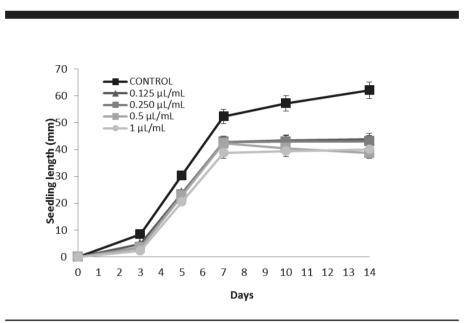


Figure 3. *L. multiflorum* seedling growth with *E. globulus* essential oil. Control and treated with *E. globulus* essential oil at 0.125, 0.25, 0.5 and 1  $\mu$ L/mL.

The herbicidal potential is closely related with the essential oil composition, weeds and doses applied. In this way, at the same doses and weed, *E. globulus* essential oil with 1,8-cineole (76.43 %) has no phytotoxic effect against *P. oleracea* seed germination and seedling growth whereas *E. camaldulensis* essential oil with spathulenol (41.46 %) as main compound was able to completely inhibit the seed germination of this cosmopolitan weed [32].

Regarding the weed, *E. globulus* essential oil showed at the doses employed significant effect on *E. crus-galli* seedling growth without phytotoxic effects against *E. crus-galli* seed germination while at higher doses (100 and 250  $\mu$ g/m) *E. tereticornis* essential oil with  $\alpha$ -pinene, 1,8-cineole and  $\beta$ -pinene as the main compounds significantly affected both seed germination and seedling growth of *E. crus-galli* [37]. Finally, lower doses of *E. globulus* essential oil showed significant effects on seed germination and seedling development of other weeds such as *A. blitoides* and *C. dactylon* [26].

### **CONCLUSION**

*E. globulus* essential oil with 1,8-cineole (76.43 %) and  $\alpha$ -pinene (14.64 %) as the main compounds represents a potentially effective bioherbicide in the management of *E. crus-galli*. Further



studies are needed with higher doses of *E. globulus* essential oil in order to corroborate a potential use also as a pre-emergent herbicide as well as its phytotoxic effects against food crops, mainly rice and other cereals.

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#### REFERENCES

- [1] Castro JA, Brasileiro BP, Lyra DH, De Almeida D. Ethnobotanical study of traditional uses of medicinal plants: The flora of caatinga in the community of Cravolândia-BA. Brazil. 2011;5(10):1905-17.
- [2] Rigat M, Vallès J, Iglésias J, Garnatje T. Traditional and alternative natural therapeutic products used in the treatment of respiratory tract infectious diseases in the eastern Catalan Pyrenees (Iberian Peninsula). J Ethnopharmacol. 2013;148(2):411-22.
- [3] Quezada RS. Therapeutic oil composition. United States Patent. Patent No.: 6,582,736 B2. 2003; June 24:1-5.
- [4] López AJ, Miranda M, Bello A, García G. Expectorant and toxicological activity of a formulation made from *Eucalyptus globulus* Labill, *Borago officinalis* L., and *Sambucus nigra* L. Rev Cuba Plantas Med. 2016;21(4):1-9.
- [5] Avello M, Fernández P, Fernández M, Schulz B, de Diego M, Mennickent S *et al.* Efecto pediculicida de una formulación en base a *Eucalyptus globulus* L. Rev Chil Infectol. 2016;33(4):433-7.
- [6] Papachristos DP, Stamopoulos DC. Toxicity of vapours of three essential oils to the immature stages of *Acanthoscelides obtectus* (Say) (Coleoptera: Bruchidae). J Stored Prod Res. 2002;38(4):365-73.
- [7] Batish DR, Singh HP, Kohli RK, Kaur S. *Eucalyptus* essential oil as a natural pesticide. For Ecol Manage. 2008;256(12):2166-74.
- [8] Kumar P, Mishra S, Malik A, Satya S. Compositional analysis and insecticidal activity of *Eucalyptus globulus* (family: Myrtaceae) essential oil against housefly (*Musca domestica*). Acta Trop. 2012;122(2):212-8.
- [9] Tyagi AK, Malik A. Antimicrobial potential and chemical composition of *Eucalyptus globulus* oil in liquid and vapour phase against food spoilage microorganisms. Food Chem. 2011;126(1):228-35.
- [10] Vilela GR, de Almeida GS, D'Arce MABR, Moraes MHD, Brito JO, da Silva MF das GF *et al.* Activity of essential oil and its major compound, 1,8-cineole, from *Eucalyptus globulus* Labill., against the storage fungi *Aspergillus flavus* Link and *Aspergillus parasiticus* Speare. J Stored Prod Res. 2009;45(2):108-11.



- [11] López-Meneses AK, Plascencia-Jatomea M, Lizardi-Mendoza J, Rosas-Burgos EC, Luque-Alcaraz AG, Cortez-Rocha MO. Antifungal and antimycotoxigenic activity of essential oils from *Eucalyptus globulus*, *Thymus capitatus* and *Schinus molle*. Food Sci Technol. 2015;35(4):664-71.
- [12] Hafsa J, Ali Smach M, Ben Khedher MR, Charfeddine B, Limem K, Majdoub H *et al.* Physical, antioxidant and antimicrobial properties of chitosan films containing *Eucalyptus globulus* essential oil. LWT Food Sci Technol. 2016;68:356-64.
- [13] Isman MB, Miresmailli S, MacHial C. Commercial opportunities for pesticides based on plant essential oils in agriculture, industry and consumer products. Phytochem Rev. 2011;10(2):197-204.
- [14] FAO. AGP Pest and Pesticide Management [Internet]. 2018 [cited 2018 Jan 7]. Available from: http://www.fao.org/agriculture/crops/thematic-sitemap/theme/pests/en/.
- [15] Abrol DP, Burgess M, Chandran RS, Clark B, Culliney TW, Feola G, et al. Integrated Pest Management. Pimentel D, Peshin R, editors. Springer; 2014. 474.
- [16] World Health Organization. Pesticide residues in food [Internet]. 2018 [cited 2018 Jan 7]. Available from: http://www.who.int/mediacentre/factsheets/pesticide-residues-food/en/.
- [17] Nandula VK, Tehranchian P, Bond JA, Norsworthy JK, Eubank TW. Glyphosate resistance in common ragweed (*Ambrosia artemisiifolia* L.) from Mississippi, USA. Weed Biol Manag. 2017;17(1):45-53.
- [18] Powles SB, Lorraine-Colwill DF, Dellow JJ, Preston C. Evolved resistance to glyphosate in rigid ryegrass (*Lolium rigidum*) in Australia. Weed Sci. 1998;46(5):604-7.
- [19] Bagavathiannan MV, Norsworthy JK, Smith KL, Neve P. Modeling the evolution of glyphosate resistance in barnyardgrass (*Echinochloa crus-galli*) in cotton-based production systems of the midsouthern United States. Weed Technol. 2013;27(3):475-87.
- [20] Synowiec A, Kalemba D, Drozdek E, Bocianowski J. Phytotoxic potential of essential oils from temperate climate plants against the germination of selected weeds and crops. J Pest Sci. 2017;90(1):407-19.
- [21] Fagodia SK, Singh HP, Batish DR, Kohli RK. Phytotoxicity and cytotoxicity of *Citrus aurantii-folia* essential oil and its major constituents: Limonene and citral. Ind Crops Prod. 2017;108:708-15.
- [22] Tworkoski T. Herbicide effects of essential oils. Weed Sci. 2002;50(4):425-31.
- [23] Hazrati H, Saharkhiz MJ, Niakousari M, Moein M. Natural herbicide activity of *Satureja hortensis* L. essential oil nanoemulsion on the seed germination and morphophysiological features of two important weed species. Ecotoxicol Environ Saf. 2017;142:423-30.
- [24] Azizi M, Fuji Y. Allelopathic effect of some medicinal plant substances on seed germination of *Amaranthus retroflexus* and *Portulaca oleraceae*. Acta Hortic. 2006;699:61-7.
- [25] Kohli RK, Batish DR, Singh HP. Eucalypt oils for the control of *Parthenium (Parthenium hysterophorus* L.). Crop Prot. 1998;17(2):119-22.
- [26] Rassaeifar M, Hosseini N, Haji Hasani Asl N, Zandi P, Moradi Aghdam A. Allelopathic effect of *Eucalyptus globulus*' essential oil on seed germination and seedling establishment of *Amaranthus blitoides* and *Cydon dactylon*. Trakia J Sci. 2013;11(1):73-81.



- [27] Adams RP. Identification of essential oil components by gas chromatography/mass spectrometry. 4th ed. Carol Stream: Allured Publishing Corporation. 2007.
- [28] Blázquez MA, Carbó E. Control of *Portulaca oleracea* by boldo and lemon essential oils in different soils. Ind Crops Prod. 2015;76:515-21.
- [29] Vieira M, Bessa LJ, Martins MR, Arantes S, Teixeira APS, Mendes *et al.* Chemical composition, antibacterial, antibiofilm and synergistic properties of essential oils from *Eucalyptus globulus* Labill. and seven Mediterranean aromatic plants. Chem Biodivers. 2017;14(6):1-12.
- [30] Harkat-Madouri L, Asma B, Madani K, Bey-Ould Z, Rigou P, Grenier D *et al.* Chemical composition, antibacterial and antioxidant activities of essential oil of *Eucalyptus globulus* from Algeria. Ind Crops Prod. 2015;78:148-53.
- [31] Ghaffar A, Yameen M, Kiran S, Kamal S, Jalal F, Munir B *et al.* Chemical composition and *in-vitro* evaluation of the antimicrobial and antioxidant activities of essential oils extracted from seven *Eucalyptus* species. Molecules. 2015;20(11):20487-98.
- [32] Verdeguer M, Blázquez MA, Boira H. Phytotoxic effects of *Lantana camara*, *Eucalyptus camaldulensis* and *Eriocephalus africanus* essential oils in weeds of Mediterranean summer crops. Biochem Syst Ecol. 2009;37(4):362-9.
- [33] Mulyaningsih S, Sporer F, Zimmermann S, Reichling J, Wink M. Synergistic properties of the terpenoids aromadendrene and 1,8-cineole from the essential oil of *Eucalyptus globulus* against antibiotic-susceptible and antibiotic-resistant pathogens. Phytomedicine. 2010;17(13):1061-6.
- [34] Hendry ER, Worthington T, Conway BR, Lambert PA. Antimicrobial efficacy of eucalyptus oil and 1,8-cineole alone and in combination with chlorhexidine digluconate against microorganisms grown in planktonic and biofilm cultures. J Antimicrob Chemother. 2009;64(6):1219-25.
- [35] Simsek M, Duman R. Investigation of effect of 1,8-cineole on antimicrobial activity of chlor-hexidine gluconate. Pharmacognosy Res. 2017;9(3):234-7.
- [36] Porres-Martínez M, González-Burgos E, Carretero ME, Pilar Gómez-Serranillos M. *In vitro* neuroprotective potential of the monoterpenes α-pinene and 1,8-cineole against H<sub>2</sub>O<sub>2</sub>-induced oxidative stress in PC12 cells. Zeitschrift fur Naturforsch Sect C J Biosci. 2016;71(7-8):191-9.
- [37] Vishwakarma GS, Mittal S. Bioherbicidal potential of essential oil from leaves of *Eucalyptus tereticornis* against *Echinochloa crus galli* L. J Biopestic. 2014;5(1):47-53.

