

Genotoxicity analysis by presence of arsenic in soil: test *Tradescantia* micronucleus extracts by clone 4430 (trad-mcn)

Análisis de genotoxicidad por presencia de arsénico en el suelo: prueba de micronucleos en extractos de *Tradescantia* clone 4430 (trad-mcn)

Análise de genotoxicidade por presença de arsênio em solo: teste de extractos de micronúcleos de *Tradescantia* pelo clone 4430 (trad-mcn)

MIGUEL ÁNGEL RICO RODRÍGUEZ¹, FRANCISCO PRIETO-GARCÍA²,
ELENA MARÍA OTAZO-SÁNCHEZ², JUDITH PRIETO MÉNDEZ³,
OTILIO ARTURO ACEVEDO-SANDOVAL³

ABSTRACT

Arsenic is a toxic metalloid that is present in air, water and soil. Soil bacteria play an important role in the oxidation of this element. This article describes the use of plants

Recibido para evaluación: 5 de Mayo de 2018.

Aprobado para publicación: 2 de Octubre de 2018.

- 1 Academic Studies Center on Environmental Pollution, University of Queretaro, Ingeniero Químico Ambiental, Cerro de las Campanas s/n. Querétaro, México.
- 2 Academic Area of Chemistry, Autonomous University of Hidalgo State. Pachuca, México.
- 3 Institute of Agricultural Sciences, Autonomous University of Hidalgo State, Rancho Universitario. Tulancingo, México.

Correspondence: prietog@uaeh.edu.mx

as bioassay for the detection of genotoxicity induced leaching of soils with arsenic (As). Test micronuclei induction was used stem cells pollen *Tradescantia* clone 4430 (Trad-MCN). The results demonstrate that the micronuclei frequency was higher for soil leachate from the community of the Salitre, Hidalgo, and the community of Bella Vista del Río, Querétaro, in Mexico. Compared with the control group of leachates soil for the Municipality of Querétaro. In all cases there was a significant difference (Tukey $p > 0,05$). Also resulting in good linear correspondence $R^2 = 0,94$ and the slope of the equation that represents a velocity induced MNs 0,5/100 tetrads each $\mu\text{gAs}\cdot\text{L}^{-1}$.

RESUMEN

El arsénico es un metaloide tóxico que está presente en el aire, el agua y el suelo. Las bacterias del suelo juegan un papel importante en la oxidación de este elemento. Este artículo describe el uso de plantas como bioensayo para la detección de lixiviación inducida por genotoxicidad de suelos con arsénico (As). Se utilizó como prueba, la inducción de micronúcleos en células de polen de *Tradescantia* clon 4430 (Trad-MCN). Los resultados demuestran que la frecuencia de aparición de micronúcleos fue más alta para el lixiviado del suelo de la comunidad de Salitre, Hidalgo y para la comunidad de Bella Vista del Río, Querétaro, en México, comparados con el grupo control de lixivados del suelo para el Municipio de Querétaro. En todos los casos hubo una diferencia significativa (Tukey $p > 0,05$). También resultó en una buena correspondencia lineal $R^2 = 0,94$ y la pendiente de la ecuación que representa una velocidad de MNs inducidos 0,05/100 tétradas cada $\mu\text{gAs}\cdot\text{L}^{-1}$.

RESUMO

O arsênico é um metalóide que está presente no ar, a água e o solo. As bactérias do solo são um importante papel na oxidação deste elemento. Este artigo descreve o uso de plantas como bioensaio para a detecção de lixiviação induzida pela genotoxicidade de suelos con arsénico (As). Use, como resultado, a indução de micronúcleos em células de pólen de *Tradescantia* clon 4430 (Trad-MCN). Resultados demuestran frecuencia de aparección de micronúcleo fue más alta para o lixiviado do suelo da comunidade de Salitre, Hidalgo y para a comunidade de Bella Vista del Río, Querétaro, en México, comparados con el grupo controle de lixivados del suelo par Municipio de Querétaro. En todos os casos principais e uma diferença significativa (Tukey $p > 0,05$). Mais resultados em uma correspondência linear $R^2 = 0,94$ e o progresso da equação que representa uma velocidade de MNs inducidos 0,05/100 tétradas cada $\mu\text{gAs}\cdot\text{L}^{-1}$.

INTRODUCTION

Arsenic is a toxic metalloid that is present in air, water and soil. In nature, it can be found as an inorganic form when combined with oxygen, sulfur and chlorides. Soil bacteria play an important role in the oxidation of this element. It is most prevalent form found in air samples is arsenic trioxi-

KEY WORDS:

Tradescantia, Micronuclei, Zimapán, Arsenic, Genotoxicity.

PALABRAS CLAVE:

Tradescantia, Micronúcleos, Zimapán, Arsénico, Genotoxicidad.

PALAVRAS-CHAVE:

Tradescantia, micronúcleos, Zimapán, Arsênico, Genotoxicidade.

de (As_2O_3) whereas a variety of arsenates (AsO_4^{3-}) or arsenite (AsO_2^-) are found in water, soil, or food [1,2]. Arsenic can act as genotoxic agent, causing adverse effects on human health and other organisms [3]. It is through inhalation of powders, ingestion of plants that have absorbed arsenic from soil, or from drinking Arsenic dissolved in that water that this element enters the food chain causing problems such as skin cancer (keratosis) as well as lung or liver cancer [4,5]. A number of studies have been conducted to detect genotoxic activity as a result of human activity, especially when mining is involved [6]. There are arsenic compounds that are known to induce mutagenic, carcinogenic and teratogenic effects resulting in a high incidence of oncological diseases [7]. Through experimental models genotoxic effects have been found in organisms exposed to these compounds. Clinical signs of these effects include gastrointestinal irritation, vomiting, diarrhea, nausea, and abdominal pain. These symptoms were observed in all cases, which were exposed to high doses of inorganic arsenic for a short-term and in low doses over a long-term [8,9]. In animal studies, lesions were found that caused gastrointestinal bleeding. For example, in monkeys that were given $6 \text{ mg As kg}^{-1} \cdot \text{day}^{-1}$ diet for a month, intestinal inflammation and bleeding were found [6]. Currently, there are a large number of bioassays both *in vitro* and as well as short and long term, to determine the genotoxicity of inorganic arsenic. *Tradescantia* clone 4430 (Hybrid X T. T. *Subcaulis hirsutiflora*) is one of the bioassays that has been used to detect genotoxic effects of heavy metals [10,11]. Many studies mention that these plants are highly sensitive to toxins found in the environment such as pesticides and other inorganic compounds [11-17]. Chemicals such as lead tetra-acetate and arsenic trioxide induced micronucleus formation in *Tradescantia* clone 4430 (Trad-MCN) [13,18]. The aim of this study was to compare the effects of genotoxicity of six leachate soils from three separate sites semi-desert states located in the states of Querétaro and Hidalgo, Mexico, with known arsenic, using a bioassay (Trad-MCN).

METHOD

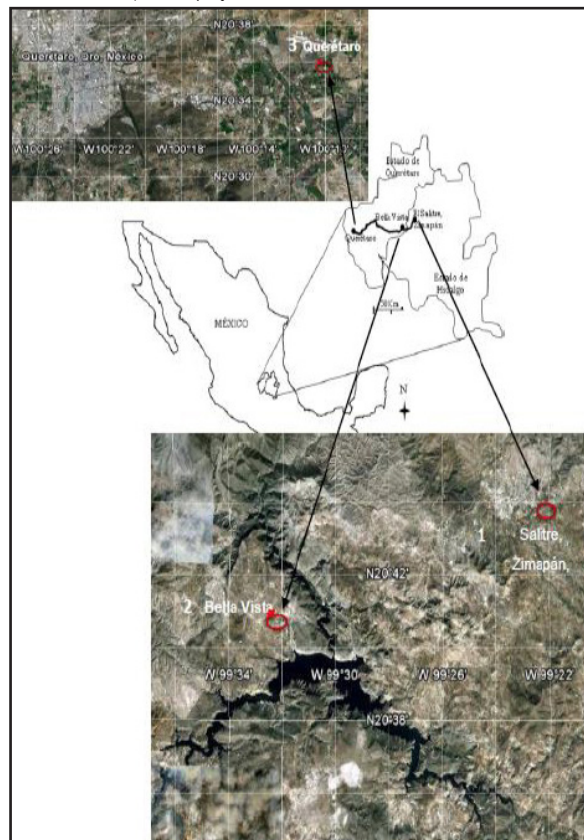
Samples

Soil and water samples from three sites located in the semi-desert area of the states of Querétaro and Hidalgo were taken. Site (1) El Salitre, is located within the

mining district of Zimapán, with an altitude of 1,780 meters above sea level and $20^\circ 44' \text{ N } 99^\circ 23' \text{ W}$; Site (2), Bella Vista del Rio which is located at $20^\circ 40' \text{ N}$ and $99^\circ 32' \text{ W}$ to 20 miles from Salitre, along the border of the states of Hidalgo and Querétaro. The third site (3), which was used as control, near the town of Querétaro $20^\circ 35' \text{ N}$ and $100^\circ 11' \text{ W}$ (Figure 1).

Pots (30 in total) were prepared with 1,5 kg of soil. The soils were washed with 500 mL of water each site; Salitre in Zimapán, Bella Vista del Rio in Querétaro, and Querétaro soils (negative control). In addition, soil from Salitre was washed with ground water from Querétaro to which a concentration of arsenic ($100 \text{ mg} \cdot \text{L}^{-1}$) was added to serve as a positive control. Over a time period of approximately four months four separate samples were obtained each with an interval of 21 days between preparations. The leachates were collected in plastic bags and kept refrigerated at -4°C until used for genotoxicity tests.

Figure 1. Location of the sampling sites. 1) Salitre, Zimapán $20^\circ 44' 51'' \text{ N } 99^\circ 22' 15'' \text{ W}$; 2) Bella Vista $20^\circ 40' 35'' \text{ N } 99^\circ 32' 16'' \text{ W}$; and 3) Querétaro $20^\circ 35' 52'' \text{ N } 100^\circ 10' 33'' \text{ W}$.



Analysis of water

Water analysis was conducted in Salitre, Zimapán, Bella Vista and Querétaro. This same water was used for watering twice a month. The pH, conductivity (EC), redox potential (Eh), and the concentration of arsenic (As) were determined. These analyzes were used as a baseline for watering the pots, following the provisions of current Official Mexican Regulations [19].

Experimental conditions for obtaining leachate

In order to collect leachate from the pots previously mentioned the following experiment was performed; Experiment A: SZ-soil Zimapán (Salitre), washed with water from WZ-Zimapán. Experiment B: (SZ)-soil Zimapán (Salitre), washed with WQ-water from Querétaro. Experiment C: (SBV)- Bella Vista, Querétaro soil, washed with WBV-Agua Bella Vista, Querétaro; Experiment D: SBV- Bella Vista, Querétaro soil, washed with WQ- water from Querétaro.

Experiment F: SQ- Queretaro soil, uncontaminated, washed with WQ-Queretaro water as a negative control. A completely randomized design was carried out, taking random samples (five replications in each case).

Selecting cuts *Tradescantia*

Tradescantia clone 4430 (that is a hybrid of *T. hirsutiflora*, X *T. subacaulis*) is heterozygous was obtained from the greenhouse of the Centro de Estudios Académicos sobre Contaminación Ambiental (CEACA) of the Autonomous University of Querétaro. Cultivation was carried out in the greenhouse under controlled conditions with light (1,800 watts Fluorescent, incandescent 180 watts) photoperiod (16/8 h light/dark), temperature 21°C and 15°C and approximately 16°C at night. The relative humidity was maintained between 60 and 80% [20].

Trad-MCN

The following protocols were used [21-23], where inflorescences were selected with an approximate stem height of 10 cm, these were immersed in the aqueous extracts for each of the six experiments. Approximately 10 sections for each experiment were used and the treatments were conducted in a greenhouse under controlled conditions. The exposure time was 6

hours with 24 hours of recovery in Hoagland nutrient solution for each sample. The inflorescences were fixed in an ethanol - acetic acid (3:1) and stained with 0,5% acetocarmine. The number of cells in tetrad stage containing 1, 2, 3 or more micronuclei and 300 cells were counted for each preparation were recorded.

Statistical Analysis

All data was analyzed using (ANOVA) variance and Tukey test ($p < 0,05$) with 5.01 JMP statistical package.

RESULTS

Characterization of the three soils and the water

The pH values, redox potential (Eh), electrical conductivity (EC) of the three soils showed some differences. The pH values classified as neutral for Querétaro soils [24,25] soils of Bella Vista and El Salitre moderately alkaline, the table 1 shows these results.

The CE of the three soils demonstrated similar and redox potential (Eh), they were classified as intermediate reducers without significant differences however with the Querétaro soil. There were significant differences [24]. Compared to Zimapán, Salitre soils where the concentrations were $(726,71 \pm 42,91 \mu\text{g As.kg}^{-1})$. These concentrations were the highest found. This is due to geomorphological origin. Next, in order of decreasing natural contamination, Bella Vista soil, with $73,25 \pm 1,35 \mu\text{g As.kg}^{-1}$, which is ten times less than the soil of Salitre. In these two cases, the concentrations indicate the geographical proximity and identify them as soils with low As concentrations ($<40 \mu\text{g As.kg}^{-1}$). The Querétaro soil which was taken only as reference contained concentrations of $12,67 \pm 1,15 \mu\text{g As.kg}^{-1}$.

Table 1. Results of the averages of analysis of the soils of the three regions studied. In parenthesis is the standard deviation.

	Site		
	Salitre, Zimapán Hidalgo	Bella Vista, Querétaro	Querétaro, Querétaro
pH	8,11 (0,06) ^a	8,18 (0,01) ^a	7,10 (0,05) ^b
[As] ($\mu\text{gAs.kg}^{-1}$)	726,71 (42,91) ^a	73,25 (1,35) ^b	12,67 (1,15) ^c
Different letters in rows indicate significant Differences ($p < 0,05$)			

The results of the characterization of the water used for irrigation are shown in cuadro 2.

A significant difference was determined in concentrations of As (in all cases HAsO_4^{-2} shaped, according to Pourbaix diagram Eh vs pH). It is observed that the initial concentration of As the water used to irrigate Zimapán exceeds 9,5 times the NOM-127-SSA1, so it is also valid for positive control; Water Bella Vista exceeds 2,5 times the standard and meets Querétaro water content $<10 \mu\text{g.L}^{-1}$.

Genotoxic activity of leachate (Tradescantia clone 4430)

There are precedents that mention that soils may contain genotoxic compounds such as the study by [17] found an increase in the frequency of micronuclei in Tradescantia clone 4430 to expose this plant outage leached from soils contaminated with heavy metals. The results for genotoxicity of leachate from soils under study, can be seen in cuadro 3. The leachate SQ/WQ was taken as negative control, because in the water of this site concentration (As) is below the allowable limit set by the NOM-127-SSA1-1994 and presented a value of $1,6 \pm 0,34$ micronuclei. The leaching of SQ/WQC with $8,5 \pm 0,37$ micronuclei taken as a positive control, showed a significant difference from the control. This difference shown in the frequency of micronuclei is attributed to the concentration of arsenate added intentionally, but unlike the results found in leachate SZ/WZ and SZ/WQ which were $7,1 \pm 0,21$ and $4,8 \pm 0,33$ micronuclei micronuclei, respectively and also showed significant difference ($p < 0,01$), the first very similar to SQ/WQC, but for this experiment also provided

soil arsenic concentration in this case in previous studies was the soil with the highest concentration ($726,71 \text{ g. kg}^{-1} \text{ As}$) and because of this it can be said that the leachate more damage caused was to SZ/WZ, corresponding with the analysis in a study by the Institute of Geophysics of the National Autonomous University of Mexico (IGEF), arsenic found in water, soil and mineral sediments, in the Zimapán population of Hidalgo.

Likewise Prieto *et al.* [26] found that the frequency of MNs in experimental Zimapán water samples showed significant differences from control in apical meristem cells of *Vicia faba*. Finally, in leachates from soils of Bella Vista SBV/WBV, it was found that this presented genotoxic damage with a frequency of $5,9 \pm 0,80$ micronuclei and SBV/WQ with a value of $5,2 \pm 0,57$ micronuclei between these two leachate no significant difference $P > 0,05$ was presented, but if compared to the control group SQ/WQ, $P < 0,01$. The values of arsenic ($73,25 \mu\text{g.kg}^{-1}$) for this site were ten times lower compared

Table 2. Average results of characterization of the water used for irrigation.

Water	pH	CE (dS/cm)	As ($\mu\text{g.L}^{-1}$) initial	As ($\mu\text{g.L}^{-1}$) Leachate
Salitre, Zimapán, Hidalgo	7,86 (0,03) ^a	0,21 (0,06) ^c	94,6 (0,18) ^d	107,32 (0,38) ^e
Bella Vista, Querétaro	7,75 (0,04) ^a	0,18 (0,04) ^c	24,6 (0,11) ^e	31,52 (0,27) ^h
Querétaro, Querétaro	8,01 (0,04) ^a	0,23 (0,06) ^c	7,2 (0,06) ^f	7,34 (0,07) ⁱ

Different letters in the columns indicate significant differences ($p < 0,05$)

Table 3. Results of the frequency of micronucle/100 tetrads, exposed soil leachate Querétaro, Queretaro, Salitre Hidalgo and Bella Vista Qro.

Leachate	Stage I		Stage II		Stage III		Stage IV		Average	
	As	MCNs	As	MCNs	As	MCNs	As	MCNs	As	MCNs
SZ/WZ	128,96	7,1	141,41	10,70	124,04	9,20	116,93	7,2	127,84±5,34	8,55±2,93
SZ/WQ	90,24	4,8	128,62	7,00	93,69	7,50	96,33	6,4	102,22±4,98	6,43±0,37
SBV/WBV	48,75	5,9	94,33	7,00	95,39	6,70	87,72	6,6	81,55±4,71	6,55±0,76
SBV/WQ	33,08	5,2	88,77	5,20	90,09	5,40	61,74	6,0	68,42±5,27	5,45±0,44
SQ/WQC(+)	89,25	4,5	150,19	9,50	151,59	8,90	156,63	8,0	136,92±5,23	7,73±0,59
SQ/WQ(-)	5,72	1,6	4,5	1,20	3,39	1,30	4,29	1,5	4,48±0,80	1,4±0,18

(+) negative control; (-) Positive Control; As in $\mu\text{g.As.kg}^{-1}$; MCNs / 100 tetrads

to that found in soil Salitre, Zimapán in Hidalgo. With the average values of the four stages the correlation between the concentration of [As] and induction of micronuclei (MNs/100 tetrads) was established. In Figure 2 the linear correspondence with an $R^2 = 0,94$ is shown, the slope of the equation (0,05) represents the speed of MCNs induced/100 tetrads per $\mu\text{gAs.L}^{-1}$. The value of the intercept represents the number of MCNs induced/100 tetrads spontaneously (see figure 3).

There are a number of studies focused on the genotoxic evaluation of soil samples, obtaining good results, focusing on the preparation of leachate and make presentations with different bioassays [27-29]. Therefore it is said that mutagenicity bioassay Proven measures to assess the genotoxicity of complex mixtures without the need to specify the chemical characterization of the sample, determining the genetic damage in living organisms [27-31].

Figure 2. Linear correlation between MNs / 100 tetrads vs [As] in $\mu\text{gAs.kg}^{-1}$.

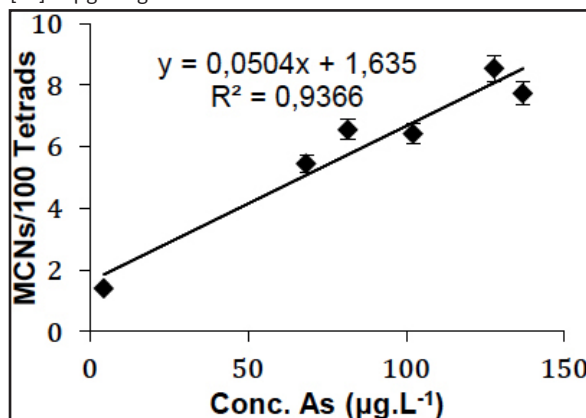
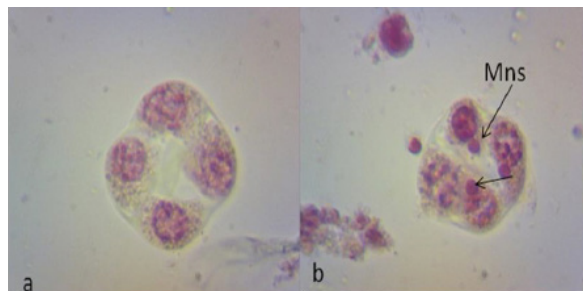


Figure 3. a) Shows a photograph of normal tetrad b) MCN in tetrads.



CONCLUSIONS

The results demonstrate that the micronuclei frequency was higher for soil leachate from the community of the Salitre, Hidalgo, and the community of Bella Vista del Río, Queretaro. Compared with the control group of leachates soil for the Municipality of Querétaro. In all cases there was a significant difference (Tukey $p > 0,05$). Also resulting in good linear correspondence $R^2 = 0,94$ and the slope of the equation that represents a velocity induced MNs 100/0.05 tetrads each $\mu\text{gAs.L}^{-1}$.

REFERENCES

- [1] GHULAM-ABBAS, B.M., IRSHAD-BIBI, M.S., NABEL-KHAN-NIAZI-ID, M.I.K., MUHAMMAD-AMJAD, M.H. and NATASHA-MAGALHAES, M.C.F. Arsenic Uptake, Toxicity, Detoxification, and Speciation in Plants: Physiological, Biochemical, and Molecular Aspects. *International Journal of Environmental Research. Public Health*, 15(59), 2018, p. 1-45, DOI:10.3390/ijerph15010059.
- [2] PARAMITA, M. An insight of environmental contamination of arsenic on animal health. *Emerging Contaminants*, 3, 2017, p. 17-22, DOI:10.1016/j.emcon.2017.01.0042405-650/.
- [3] SEO, M.N., LEE, S.G. and EOM, S.Y. Estimation of total and inorganic arsenic intake from the diet in Korean adults. *Archives of Environmental Contamination and Toxicology*, 70, 2016, p. 647-656.
- [4] XUESONG, X.I., LU, L., CHARALAMBOS, P. and PEI, X. Sorption of Arsenic from Desalination Concentrate onto Drinking Water Treatment Solids: Operating Conditions and Kinetics. *Water*, 10(96), 2018, p. 1-15, DOI:10.3390/w10020096.
- [5] BAE, H.S., KANG, I.G., LEE, S.G., EOM, S.Y., KIM, Y.D., OH, S.Y., KWON, H.J., PARK, K.S., KIM, H., CHOI, B.S., YU, I.J. and PARK, J.D. Arsenic exposure and seafood intake in Korean adults. *Human and Experimental Toxicology*, 36(5), 2017, p. 451-460.
- [6] BAIG, S.A., SHENG, T., HU, Y., XU, J. and XU, X. Arsenic removal from natural water using low cost granulated adsorbents: A review. *CLEAN Soil Air Water*, 43, 2015, p. 13-26.
- [7] GORBOV, S.N., BEZUGLOVA, O.S., VARDUNI, T.V., GOROVTSOV, A.V., TAGIVERDIEV, S.S. and HILDEBRANT, Y.A. Genotoxicity and Contamination of Natural and Anthropogenically Transformed Soils of the City of RostovonDon with Heavy Metals. *Eurasian Soil Science*, 48(12), 2015, p. 1383-1392.

- [8] HABUDA-STANI, M., NUJIC, M., ROMIC, Ž., LONCARIC, A., RAVANCIC, M.E. and KRALJ, E. Arsenic preoxidation and its removal from groundwater using iron coagulants. *Desalination Water Treatment*, 56, 2015, p. 2105–2113.
- [9] ABEJÓN, A., GAREA, A. and IRABIEN, A. Arsenic removal from drinking water by reverse osmosis: Minimization of costs and energy consumption. *Separation and Purification Technology*, 144, 2015, p. 46–53.
- [10] TE-HSIU, M.A., XU, Z., XU, C., MCCONNELL, H., RABAGO, E.V., ARREOLA, G.A. and ZHANG, H. The improved Allium/Vicia root tip micronucleus assay for clastogenicity of environmental pollutants. *Mutation research*, 334(2), 1995, p. 185–95.
- [11] MELO-DE-CARVALHO, R., LAÍS -COUTO-MACHADO, J., SOUSA DE AGUIAR, R.P., OLIVEIRA-FERREIRA DA-MATA, A.M., RODRIGUES-SILVA, R., SILVA-TEIXEIRA, J., OLIVEIRA-BARROS-DE-ALENCAR, M.V., TOREQUL-ISLAM, M. and DE CARVALHO-MELO-CAVALCANTE, A.A. *Tradescantia pallida* as a biomonitoring tool to assess the influence of vehicle exhaust and benzene derivatives. *African Journal of Biotechnology*, 16(6), 2017, p. 280–287, DOI: 10.5897/AJB2017.15897.
- [12] SANTOS, A.P.M., SEGURA-MUÑOZ, S.I., NADAL, M., SCHUHMACHER, M. and DOMINGO J.L. Traffic-related air pollution biomonitoring with *Tradescantia pallida* (rose) Hunt. cv. purpurea Boom in Brazil. *Environmental Monitoring and Assessment*, 187(2), 39, 2015, p. 1–10.
- [13] LIMA, A.V.A., BARBOSA, M.A.S., CUNHA, L.C.S., DE MORAIS, S.A.L., DE AQUINO, F.J.T., CHANG, R. and DO NASCIMENTO, E.A. Volatile Compounds Obtained by the Hydrodistillation of Sugarcane Vinasse, a Residue from Ethanol Production. *Revista Virtual de Química*, 2017, 9(2), p. 764–773, DOI: 10.21577/1984-6835.20170047.
- [14] SANTOS, A.P., SEGURA-MUÑOZ, S.I., NADAL, M., SCHUHMACHER, M., DOMINGO, J.L., MARTINEZ, C.A. and MAGOSSO-TAKAYANAGUI, A.M. Traffic-related air pollution biomonitoring with *Tradescantia pallida* (Rose) Hunt. cv. purpurea Boom in Brazil. *Environmental Monitoring and Assessment*, 187, 2015, p. 1–10.
- [15] BETÂNIA-BRIZOLA-CASSANEGO, M., MARQUES-DA-COSTA, G., HISAYUKI-SASAMORI, M., ENDRES-JÚNIOR, D., TAMIREZ-PETRY, C. and DROSTE, A. The *Tradescantia pallida* var. purpurea active bioassay for water monitoring: evaluating and comparing methodological conditions. *Revista Ambiente y Agua*, 9(3), 2014, p. 424–433, DOI: 10.4136/ambi-agua.1411.
- [16] OLLER-CRUZ, O.J. Empleo de bioindicadores para determinar la calidad del aire en la ciudad de Tarija en puntos de muestreo de red Moni-CA. *Acta Nova*, 8(3), 2018, p. 307–321.
- [17] PO-WEN, C., ZHEN-SHU, L., MIN-JIE, W. and TAI-CHEN, K. Cellular Mutagenicity and Heavy Metal Concentrations of Leachates Extracted from the Fly and Bottom Ash Derived from Municipal Solid Waste Incineration. *International Journal of Environmental Research and Public Health*, 13(11), 2016, p. 1078, <https://doi.org/10.3390/ijerph13111078>.
- [18] MISIK, M., MA, T.H., NERSESYAN, A., MONARCA, S., KIM, J.K. and KNASMUELLER, S. Micronucleus assays with *Tradescantia* pollen tetrads: an update. *Mutagenesis*, 26(1), 2011, p. 215–221.
- [19] MÉXICO. NOM-127-SSA1-1994. Norma Oficial Mexicana Salud ambiental, agua para uso y consumo humano- Límites permisibles de calidad y tratamiento a que debe someterse para su potabilización. México D.F. (México): Diario Oficial de la Federación, 1996.
- [20] BAKARE, A.A., ALIMBA, C.G. and ALABI, O.A. Genotoxicity and mutagenicity of solid waste leachates: A review. *African Journal of Biotechnology*, 12(27), 2013, p. 4206–4220.
- [21] CURADO, A.L., CUNHA DE OLIVEIRA, C., COSTA, W.R., BORELLA-MARFIL-ANHÊ, A.C. and MILLA- DOS SANTOS-SENHUK, A.P. Urban influence on the water quality of the Uberaba River basin: an ecotoxicological assessment. *Revista Ambiente y Agua*, 13(1), 2018, p. 1–10.
- [22] CARTER, L.J., HARRIS, E., WILLIAMS, M., RYAN, J.J., KOOKANA, R.S. and BOXALL, A.L. Fate and Uptake of Pharmaceuticals in Soil–Plant Systems. *Journal of Agricultural and Food Chemistry*, 62(4), 2014, p. 816–825, DOI: 10.1021/jf404282y.
- [23] DHYEVRE, A., FOLT ETE, A.S., ARAN, D., MULLER, S. and COTELLE, S. Effects of soil pH on the Vicia-micronucleus genotoxicity assay. *Mutation Research- Genetic Toxicology and Environmental Mutagenesis*, 774, 2014, p. 17–21.
- [24] BOULDING, J.R. Description and sampling of contaminated soils. A field guide. 2 ed. eBook ISBN 9781351456142. Boca Raton (USA): FL7 Lewis Publishers, 2017, p. 12–16. file:///C:/Users/Administrador/Documents/Downloads/9781351456142_preview.pdf

- [25] MEDINA-GUERRERO, M.L. y ZANOR, G.A. Evaluación de la contaminación por elementos traza en suelos agrícolas de municipio de Ira-puato (Guanajuato). *Jóvenes en la Ciencia*, 3(1), 2017, p. 295-299.
- [26] PRIETO, G.F., LECHUGA, V.M.A., MÉNDEZ, M.M.A., BARRADO, E.E.Y. y OYARZÚN, G.J. Daños tóxicos en tejidos vegetales, producidos por aguas contaminadas con arsénico en Zimapan, Hidalgo México. *Ciência e Tecnologia de Alimentos*, 26(1), 2006, p. 94-97
- [27] CHAHAL, V., NAGPAL, A., PAKADE, Y.B. and KATNORIA, J.K. Ecotoxicological Studies of Soil Using Analytical and Biological Methods: A Review. *International Journal of Biotechnology and Bioengineering*, 8(3), 2014, p. 302-318.
- [28] MUNAWAR, I. *Vicia faba* bioassay for environmental toxicity monitoring: A review. *Chemosphere*, 144, 2016, p. 785-802.
- [29] MENESTRINO-GARCIA, E., RODRIGUES-DA SILVA-JUNIOR, F.M. and MUCCILLO-BAISCH, A.L. Mutagenic effect of contaminated soil on the offspring of exposed rats. *Acta Scientiarum, Health Sciences Maringá*, 38(1), 2016, p. 19-22.
- [30] SABZAR, A.D., ABDUL-REHMAN, Y. and MASOOD-UL-HASSAN, B. An Introduction about Genotoxicology Methods as Tools for Monitoring Aquatic Ecosystem: Present Status and Future Perspectives. *Fisheeries and Aquaculture Journal*, 7, 158, 2016, p. 1-11, DOI:10.4172/2150-3508.1000158.
- [31] RASHMI, K., SHELJA, S. and AVINASH, K.N. Comparative antigenotoxic effects of aqueous leaf extracts of different cultivars of *Chrysanthemum morifolium* R. against genotoxicity induced by mercuric chloride using *Allium cepa* L. root chromosomal aberration assay. *Journal of Innovations in Pharmaceutical and Biological Sciences*, 5(2), 2018, p. 87-92.