



***Aelurostrongylus abstrusus* infection of domestic and wild cats in south Brazil**

*Infecção por Aelurostrongylus abstrusus em gatos domésticos e selvagens no sul do Brasil*

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Artigo

**Abstract:** *Aelurostrongylus abstrusus* is a helminthic pathogen that infects the respiratory tract of felids after the ingestion of intermediate or paratenic hosts. The present paper assesses *A. abstrusus* infection in domestic and wild cats in south of Brazil. Fecal samples from 97 domestic cats and from seven wild cats (*Puma concolor*, *Puma yagouaroundi* and *Leopardus tigrinus*) were tested using sedimentation and flotation techniques. The prevalence of first-stage larvae was 2.1% (2/97) in domestic cats, and *Puma yagouaroundi* was the only wild cat that turned out to be positive for this lungworm. This is the first report of *A. abstrusus* in *Puma yagouaroundi* in Brazil. The identification of this parasite raises a red flag about the presence of infected intermediate and/or paratenic hosts and about the importance of this parasitic disease even in the absence of respiratory clinical signs. The analysis of fecal samples is then suggested for the detection of *A. abstrusus*.

**Key words:** *Aerulostrongylus abstrusus* - Cat - Wild cat - Coprological methods

**Resumo:** *Aelurostrongylus abstrusus* é um patógeno helmíntico que infecta o trato respiratório de felinos após a ingestão de hospedeiros intermediários ou paratênicos. O presente trabalho avalia a infecção por *A. abstrusus* em gatos domésticos e selvagens no sul do Brasil. Amostras fecais de 97 gatos domésticos e de sete gatos selvagens (*Puma concolor*, *Puma yagouaroundi* e *Leopardus tigrinus*) foram testadas usando técnicas de sedimentação e flotação. A prevalência de larvas de primeiro estágio foi de 2,1% (2/97) em gatos domésticos, e *Puma yagouaroundi* foi o único gato selvagem que revelou ser positivo para esse verme pulmonar. Este é o primeiro relato de *A. abstrusus* em *Puma yagouaroundi* no Brasil. A identificação desse parasita levanta uma bandeira vermelha sobre a presença de hospedeiros intermediários e / ou paratênicos infectados e sobre a importância dessa doença parasitária mesmo na ausência de sinais clínicos respiratórios. A análise de amostras fecais é então sugerida para a detecção de *A. abstrusus*. Palavras-chave: *Aerulostrongylus abstrusus* - Gato - Gato selvagem - Métodos coprológicos.

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## Introduction

*Aelurostrogylus abstrusus* (Railliet 1898) is a helminthic parasite of the respiratory tract of felids, having a complex life cycle and causing infection indirectly by the ingestion of intermediate hosts infected by molluscs of the genera *Deroceras*, *Arion*, *Agriolimax*, *Helicella*, *Limax* and *Theba* or of paratenic hosts such as birds, rodents, amphibians, and lizards carrying third-stage larvae (L3) (Nabais et al. 2013). After ingested by the definitive host, the larvae migrate through the gastrointestinal tract and bloodstream, lodge in the branches of pulmonary arteries, alveoli, alveolar ducts, and bronchioles (Ribeiro and Lima 2010), and undergo two molts before they become adults.

Thereafter, they reproduce, hatch in the alveoli and develop into first-stage larvae (L1), which are coughed up, swallowed, and eventually passed in the feces (Ellis et al. 2010). Clinical signs vary according to the severity of infection, to the age of the

animal, to immune system involvement, parasite load, and to associations with other conditions. Lungworm infections in healthy individuals are subclinical and self-limiting, but in immunocompromised animals, clinical signs include labored breathing, sneezing, cough, mucopurulent nasal discharge, dyspnea, tachypnea, and eventually lead to death, as a result of oviposition in the host's bronchi, triggering an inflammatory response and, consequently, causing granulomatous pneumonia with hyperplasia and hypertrophy of the smooth muscles of the parenchyma and pulmonary arteries (Barutzi and Schaper 2012; Traversa and Di Cesare 2013). The diagnosis of *A. abstrusus* is based on the detection of larvae (L1) in fecal samples. The sedimentation method (Hoffman et al. 1934) is the gold standard for the diagnosis of this infection (Traversa et al. 2010).

Tracheal or bronchoalveolar lavage, albeit invasive, may be used, as well as thoracic radiographs (Lacorcia

et al. 2009); serological tests, however, can show cross-reactivity between endoparasite species (Traversa and Guglielmini 2008), whereas molecular methods have been found to have higher sensitivity and specificity (Traversa et al. 2008a). The aim of this paper is to assess the prevalence of *A. abstrusus* larvae in fecal samples from domestic and wild cats in southern Brazil.

### Materials and methods

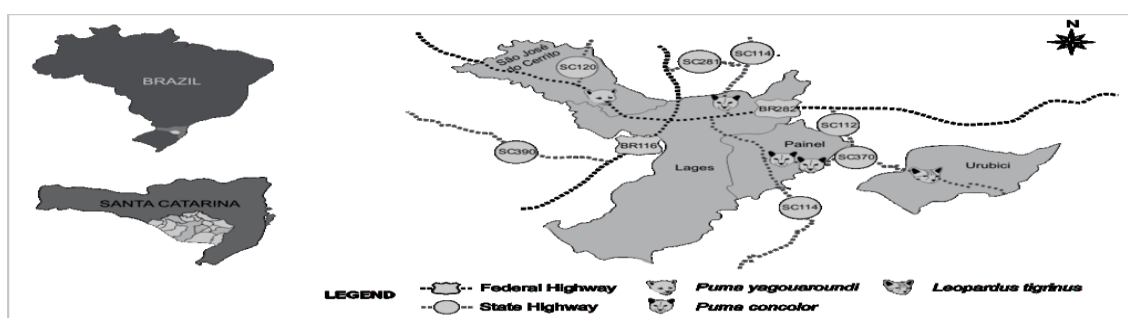
Fecal samples were collected from 97 stray cats captured from several neighborhoods in Lages (27° 49' 0" S and 50° 19' 35" W), in the mountain region of the state of Santa Catarina, southern Brazil. The cats were housed at the local Center for Zoonotic Disease Control. 24-hour stool samples were collected from the floor of individual stalls between May and November 2012.

Seven dead adult wild cats, including *Puma concolor* (3), *Leopardus tigrinus* (3) and *Puma yagouaroundi* (1), which had been

struck and killed on two highways (1 federal and 1 state), in the vicinity of Lages (Fig 1), were picked up between August 2007 and September 2012 by the Brazilian Environmental Agency (IBAMA) and sent to the Laboratory of Zoology and Parasitology of the Universidade do Planalto Catarinense (UNIPLAC).

The fecal samples were obtained directly from the wild cats' rectums at necropsy, frozen at 4°C, processed within 24 h, and analyzed by two conventional methods: sedimentation (Hoffman et al. 1934) and flotation (Faust et al. 1938). Five grams of each sample were used in each method after homogenization (Ehlers et al. 2013). The larvae (L1) were observed under light microscopy using 100X and 400X magnifications. The characteristics of L1, that is, the presence of a dorsal spine on the tail, allowed making a distinction from other nematode larvae (TRAVERSA AND DI CESARE 2013).

**Fig. 1** Highways in Lages and neighboring towns in Santa Catarina, Brazil, where roadkills took place (2007-2012)

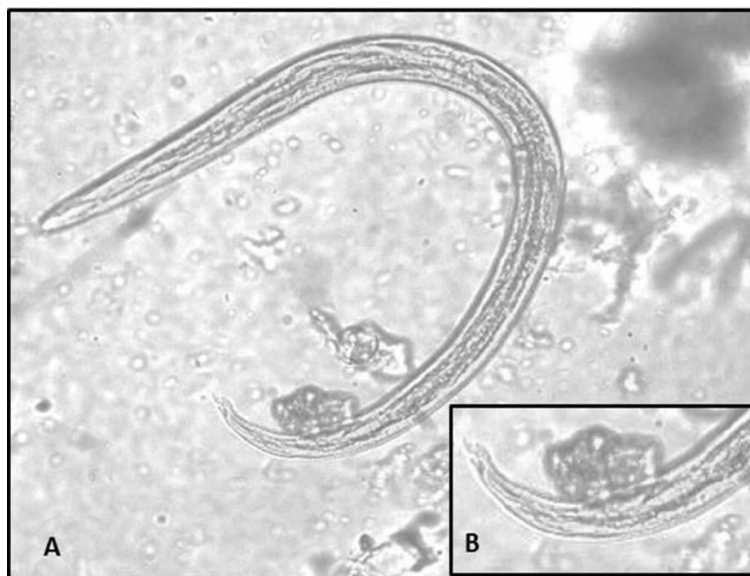


## Results

Two out of 97 stray cats, one aged 6 months and one aged four years, were positive for *A. abstrusus* L1, yielding a 2.1% rate. Among wild cats,

only *Puma yagouaroundi* was positive for this lungworm (14.3%). The larvae (Fig 2) were detected only by the sedimentation method, showing, on average, five L1 per slide.

**Fig. 2** A - *Aelurostrongylus abstrusus* larva (100X). B – Dorsal spine on the tail (400X)



## Discussion

This is the first systematic study specifically aimed at verifying the presence of *A. abstrusus* in the local stray cat population and among free-ranging wild cats, whose prevalence corresponds to 2.1% and 14.3%, respectively. *A. abstrusus* infection in domestic cats is a widely geographically distributed parasitic disease, considered to be both emergent and re-emergent. In Brazil, the prevalence rates among domestic cats range from 1.37% to 29.5% (Ohlweiler et al. 2010; Ehlers et al. 2013; Ramos et al. 2013), depending on the region.

Actually, a 2.1% prevalence rate among domestic cats is deemed to be low. The sample size, though, did not allow determining the prevalence among wild felids and, therefore, it was only possible to infer that there exist intermediate and/or paratenic hosts acting as vectors of transmission in both urban and wild areas of the investigated region. Both native and exotic mollusk species have been infected with *A. abstrusus* larvae. Around 40% of 45 specimens of *Achatina fulica* Bowdich, 1822 studied in Brazil were infected by this parasite (Andrade-Porto et al. 2012). The gastropod *A. fulica*, native to

southeastern Africa, was introduced into the American continent in 1939 and was brought to Brazil for commercial purposes in the 1980s. Nowadays, this snail is regarded as one of the most invasive pests, and is found in 23 out of 26 Brazilian states, acting an important vector of helminthic pathogens to animals and humans (Thiengo et al. 2008; Ohlweiler et al. 2010).

In Europe, most of the reports on the infection of domestic cats were published from 2000 onwards, showing different prevalence rates, depending on the cat population and on the geographic region. In Spain, the prevalence was 1% in domestic, stray, and farm cats (Miró et al. 2004); 28.7% in stray cats in Ibiza (Jefferies et al. 2010); 17.3% and 18.5% in central and southern Italy, respectively (Traversa et al. 2008b), and 2.9% in northern Italy (Spada et al. 2013); 5.6% in Romania (Mircean et al. 2010); and 14.5% in Hungary (Capári et al. 2013). In Denmark and in Germany, the prevalence rates were 5.6% (Taubert et al. 2009), but Barutzki and Schaper (2012) pointed out the disease was endemic in Germany.

*A. abstrusus* infection has been investigated all around the world: in Turkey (Tüzer et al. 2002), in Portugal (Waap et al. 2013), in Greece (Ellis et

al. 2010), in Qatar (Abu-Madi et al. 2007), in Australia (Lacorcia et al. 2009), and in the United States (Lucio-Forster and Bowman 2011); with only one case being reported in Turkey (Tüzer et al. 2002), Greece (Ellis et al. 2010), and Albania (Knaus et al. 2011). Evidence of higher prevalence rates and new foci of *A. abstrusus* infection in European countries may be the result of the larger number of intermediate hosts, possibly due to global climate changes, in addition to the fact that most cats have access to the street, where they can come in contact with fecal matter from other cats and where they can ingest intermediate and/or paratenic hosts (Traversa et al. 2010).

In Thailand, *A. abstrusus* was reported in leopards (*Panthera pardus*=55.55%), tigers (*Panthera tigris*=73.68%), and Asian leopard cats (*Felis bengalensis* =100%) (Patton and Rabinowitz 1994); in Tanzania, in lions (*Panthera leo* =7%) (Bjork et al. 2000). In Bolivia, the prevalence in wild felines (*Oncifelis geoffroyi*, *Leopardus pardalis* and *Herpailurus yaguarondi*) averaged 42.11%, corresponding to 50% (5/10) in *L. pardalis* and 37.5% (3/8) in *O. geoffroyi* (Fiorello et al. 2006). In Poland, the parasite was detected in *Lyx liyx* (21.1%) (Szczena et al. 2006), and in eastern Siberia,

González et al. (2007) detected larvae in feces of *P. tigris altaica* and *F. bengalensis euphilurus*.

*A. abstrusus* prevalence rates could be underestimated as infection may be asymptomatic and as costly and effective techniques are not always available in the routine practice of clinics and veterinary hospitals (Traversa and Guglielmini 2008; Traversa et al. 2008a; Lacorcia et al. 2009). L1 can remain viable in the feces at an ambient temperature of 21°C to 24°C for up to 45 days and at 4°C for 60 days (Gökpınar and Yildiz 2010), indicating that the appropriate disposal of feline waste plays a crucial role. Attention should be given to the life cycle and to the pathogenicity of this parasite, given that L1 are shed intermittently in the feces; hence, it is recommended that a 24-hour sample be collected or that the collection occur three times daily (Barutzi and Schaper 2012), preventing false-negative results, which are likely to be observed when the infection is still in the prepatent period (Traversa et al. 2010).

The study of wild animals presents enormous difficulties, and reports on these species are scarcely available. This study could be undertaken because it was possible to

pick up animals struck and killed by motor vehicles. These roadkills happen because urbanization has expanded hugely, thereby reducing wilderness areas, forcing felines to seek out food and sex partners elsewhere and to roam into urban spaces.

The distribution of helminthic pathogens, presence of clinical signs, physical status, epidemiological variables, and identification of intermediate and/or paratenic hosts need to be investigated. Therefore, as a suggestion, public policies should be devised, including the detection of L1 by routine parasitological tests, especially in stray domestic cats, and getting these animals off the street. The flotation method was also used, as in other studies (Gaglio et al. 2008; Traversa et al. 2008b; Lucio-Forster and Bowman 2011; Barutzi and Schaper 2012); however, it is not efficient in detecting first-stage larvae in domestic and wild cats.

The identification of *A. abstrusus* raises a red flag about the presence of infected intermediate and/or paratenic hosts and about the importance of this parasitic disease even when respiratory clinical signs are absent. This is the first report of *A. abstrusus* in *Puma yagouaroundi* in

Brazil.

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### Compliance with ethical standards

All of the studies reported herein were performed in compliance with current, applicable, local laws and regulations.

### Conflict of interest

None

### References

- ABU-MADI, M.A., AL-AHBABI, D.A., AL-MASHHADANI, M.M., AL-IBRAHIM, R., PAL, P., LEWIS, J.W., 2007. Patterns of parasitic infections in faecal samples from stray cat populations in Qatar. *Journal of Helminthology* 81, 281-286.
- ANDRADE-PORTO, S.M., SOUZA, K.C.P., CÁRDENAS, M.Q., ROQUE, R.A., PIMPÃO, D.M., ARAÚJO, C.S., MALTA, J.C.O., 2012. Occurrence of *Aelurostrongylus abstrusus* (Railliet, 1898) larvae (Nematoda: Metastrongylidae) infecting *Achatina (Lissachatina) fulica* Bowdich, 1822 (Mollusca: gastropoda) in Amazon region. *Acta Amazonica* 42, 245-250.
- BARUTZI, D., SCHAPER, R., 2012. Occurrence and regional distribution of *Aelurostrongylus abstrusus* in cats in Germany. *Parasitology Research* 112, 855-861.
- BJORK, K.E., AVERBECK, G.A., STROMBERG, B.E., 2000. Parasites and parasite stages of free-ranging wild lions (*Panthera leo*) of Northern Tanzania. *Journal of Zoo and Wildlife Medicine* 31, 56-61.
- CAPÁRI, B., HAMEL, D., VISSER, M., WINTER, R., PFISTER, K., REHBEIN, S., 2013. Parasitic infections of domestic cats, *Felis catus*, in western Hungary. *Veterinary Parasitology* 192, 33-42.
- EHLERS, A., MATTOS, M.J.T., MARQUES, S.M.T., 2013. Prevalência de *Aelurostrongylus abstrusus* (Nematoda, Strongylida) em gatos de Porto Alegre, Rio Grande do Sul. *Revista da Faculdade de Zootecnia, Veterinária e Agronomia (FZVA)* 19, 97-104.
- ELLIS, A.E., BROWN, C.A., YABSLEY, M.J., 2010. *Aelurostrongylus abstrusus* larvae in the colon of two cats. *Journal of Veterinary Diagnostic Investigation* 22, 652-655.
- FAUST, E.C., D'ANTONIO, J.S., ODOM, V., MILLER, M.J., PERES, C., SAWITZ, W., THOMEN, L.F., TOBLE, J., WALKER, J.H., 1938. A Critical study of clinical laboratory techniques for the diagnosis of protozoan cyst and helminth egg in feces. *The American Journal of Tropical Medicine and Hygiene* 18, 169-183.
- FIGLIOLLO, C.V., ROBBINS, R.G., MAFFEI, L., WADE, S.E., 2006. Parasites of free-ranging small canids and felids in the Bolivian Chaco. *Journal of Zoo and Wildlife Medicine* 37, 130-134.
- GAGLIO, G., CRINGOLI, G., RINALDI, L., BRIANTI, E., GIANNETTO, S., 2008. Use of the FLOTAC technique for the diagnosis of *Aelurostrongylus abstrusus* in the cat. *Parasitology Research* 103, 1055-1057.
- GÖKPINAR, S., YILDIZ, K., 2010. The effect of different temperatures on viability of *Aelurostrongylus abstrusus* first stage larvae in faeces of cats. *Türkiye Parazitoloji Dergisi* 34, 102-105.
- GONZÁLEZ, P., CARBONELL, E., URIOS, V., ROZHNOV, V.V., 2007. Coprology of *Panthera tigris altaica* and *Felis bengalensis euptylurus* from the Russian far east. *Journal of Parasitology* 93, 948-950.

- HOFFMAN, W.A., PONS, J.A., JANER, J.L., 1934. The sedimentation concentration method in *Schistosomiasis mansoni*. Puerto Rico Journal of Public Health 9, 283-298.
- JEFFERIES, R., VRHOVEC, M.G., WALLNER, N., CATALAN, D.R., 2010. *Aelurostrongylus abstrusus* and *Troglostrongylus* sp. (Nematode: Metastrongyloidea) infections in cats inhabiting Ibiza, Spain. Veterinary Parasitology 173, 344-348.
- KNAUS, M., KUSI, I., RAPTI, D., XHAXHIU, D., WINTER, R., VISSER, M., REHBEIN, S., 2011. Endoparasites of cats from the Tirana area and the first report on *Aelurostrongylus abstrusus* (Railliet, 1898) in Albania. Wiener Klinische Wochenschrift 123, 31-35.
- LACORCIA, L., GASSER, R.B., ANDERSON, G.A., BEVERIDGE, I., 2009. Comparison of bronchoalveolar lavage fluid examination and other diagnostic techniques with the Baermann technique for detection of naturally occurring *Aelurostrongylus abstrusus* infection in cats. Journal of the American Veterinary Medical Association 235, 43-49.
- LUCIO-FORSTER, A., BOWMAN, D.D., 2011. Prevalence of fecal-borne parasites detected by centrifugal flotation in feline samples from two shelters in upstate New York. Journal of Feline Medicine and Surgery 13, 300-304.
- MIRCEAN, V., TITILINCU, A., VASIL, C., 2010. Prevalence of endoparasites in household cat (*Felis catus*) populations from Transylvania (Romania) and association with risk factors. Veterinary Parasitology 17, 163-166.
- MIRÓ, G., MONTOYA, A., JIMÉNEZ, S., FRISUELOS, C., MATEO, M., FUENTES, I., 2004. Prevalence of antibodies to *Toxoplasma gondii* and intestinal parasites in stray, farm and household cats in Spain. Veterinary Parasitology 126, 249-255.
- NABAIS, J., ALHO, A.M., VICENTE, G., CARVALHO, L.M., 2013. Aelurostrongilose felina: uma parasitose de cortar a respiração. Vet Med Jul-ago:51-56.
- OHLWEILER, F.P., GUIMARÃES, M.C.A., TAKAHASHI, F.Y., EDUARDO, J.M., 2010. Current distribution of *Achatina fulica*, in the State of São Paulo including records of *Aelurostrongylus abstrusus* (Nematoda) larvae infestation. Revista do Instituto de Medicina Tropical 52, 211-214.
- PATTON, S., RABINOWITZ, A.R., 1994. Parasites of wild feline in Thailand: A coprological survey. Journal of Wildlife Diseases 30, 472-475.
- RAMOS, D.G.S., SCHEREMETA, R.G.A.C., OLIVEIRA, A.C.S., SINKOK, A.L., PACHECO, R.C., 2013. Survey of helminth parasites of cats from the metropolitan area of Cuibá, Mato Grosso, Brazil. Revista Brasileira de Parasitologia Veterinária 22, 201-206.
- RIBEIRO, V.M., LIMA, W.S., 2010. Larval production of cats infected and re-infected with *Aelurostrongylus abstrusus* (Nematoda: Protostrongylidae). Revue de Médecine Vétérinaire 152, 815-820.
- SPADA, E., PROVERBIO, D., DELLA PEPA, A., DOMENICHINI, G., BAGNAGATTI, DI GIORGI, G., TRALDI, G., FERRO, E., 2013. Prevalence of faecal-borne parasites in colony stray cats in northern Italy. Journal of Feline Medicine and Surgery 15, 672-677.
- SZCZESNA, J., POPIOLEK, M., SCHMIDT, K., KOWALCZYK, R., 2006. The first record of *Aelurostrongylus abstrusus* (Angiostrongylidae: Nematoda) in Eurasian lynx (*Lynx lynx* L.) from Poland based on fecal analysis. Wiadomości Parazytologiczne 52, 321-322.
- TAUBERT, A., PANTCHEV, N., VRHOVEC, M.G., BAUER, C., HERMOSILLA, C., 2009. Lungworm infections (*Angiostrongylus vasorum*, *Crenosoma vulpis*, *Aelurostrongylus abstrusus*) in dogs and cats in Germany and Denmark in 2003-2007. Veterinary Parasitology 159, 175-180.
- THIENGO, S.C., FERNANDEZ, M.A., TORRES, E.J., COELHO, P.M., LAFREDI, R.M., 2008. First record of a nematode Metatrongyliloidea (*Aelurostrongylus abstrusus* larvae) in



*Achatina fulica* (Lissachatina) *fulica* (Mollusca, Achatinidae) in Brazil. *Journal of Invertebrate Pathology* 98, 34-39.

TRAVERSA, D., DI CESARE, A., 2013. Feline lungworms: What a dilemma. *Trends in Parasitology* 29, 423-430.

TRAVERSA, D., DI CESARE, A., CONBOY, G., 2010. Canine and feline cardiopulmonary parasitic nematodes in Europe: emerging and underestimated. *Parasites & Vectors* 3, 1-22.

TRAVERSA, D., DI CESARE, A., MILILLO, P., LORIO, R., OTRANTO, D., 2008A. *Aelurostrongylus abstrusus* in a feline colony from central Italy: clinical features, diagnostic procedures and molecular characterization. *Parasitology Research* 103, 1191-1196.

TRAVERSA, D., GUGLIELMINI, C. 2008. Feline aelurostrongylosis and canine angiostrongylosis: A challenging diagnosis for two emerging verminous pneumonia infections. *Veterinary Parasitology* 157, 163-174.

TRAVERSA, D., LIA, R.P., LORIO, R., BOARI, A., PARADIES, P., CAPELLI, G., AVOLIO, S., OTRANTO, D., 2008B. Diagnosis and risk factors of *Aelurostrongylus abstrusus* in cats from Italy. *Veterinary Parasitology* 153, 182-186.

TÜZER, E., TOPARLAK, M., GARGILI, A., KELES, V., ULUTAS ESATGIL, M., 2002. A case of *Aelurostrongylus abstrusus* infection in a cat in Istanbul, Turkey and its treatment with moxidectin and levamisole. *Turkish Journal of Veterinary Animal Science* 26, 411-414.

WAAP, H., GOMES, J., NUNES, T., 2013. Parasite communities in stray cat populations from Lisbon, Portugal. *Journal of Helminthology* 30, 1-7.